



National Institutes of Health  
*The Nation's Medical Research Agency*



National Institutes of Health -- Center for Information Technology



Division of Computational Bioscience

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**Structural information  
from protein dynamics:  
restraints on protein shape encoded in  
protein rotational diffusion tensor**

## **OUTLINE**

A very general concept of protein structure elucidation:  
Local and global structural restraints.

Global restraints on protein shape from protein dynamics:  
Efficient calculations of protein diffusion tensor.

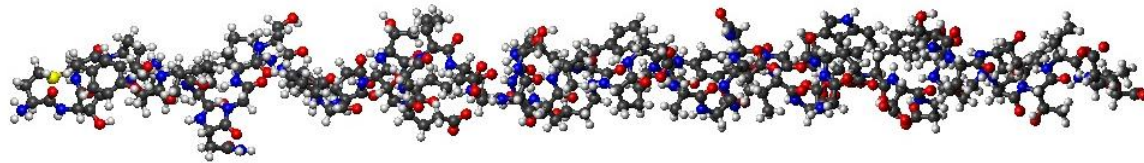
Refinement of globular protein structure using overall shape  
restraints encoded in protein diffusion tensor.

Docking domains in protein complexes.

Future directions.

# Ultimate Goal of protein structure prediction

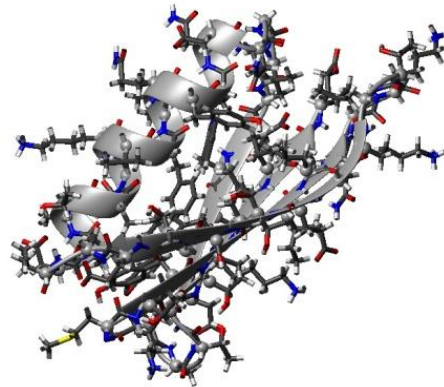
Sequence of amino acid residues



+

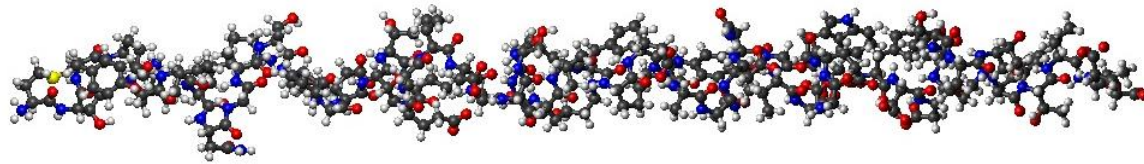
Model inter atomic forces

=

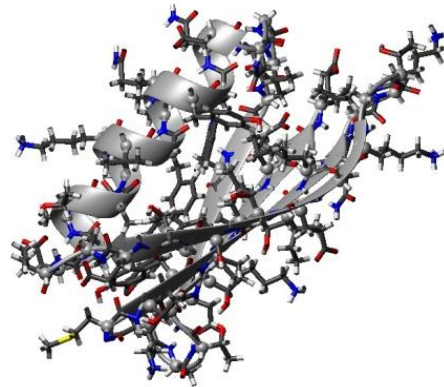


# Ultimate Goal of protein structure prediction

Sequence of amino acid residues



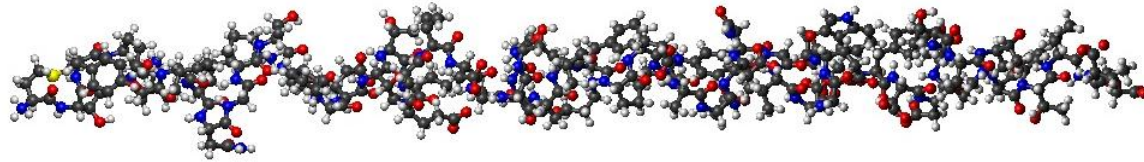
Model inter atomic forces



**Is not yet  
accomplished**

# NMR Protein structure determination

Sequence of amino acid residues



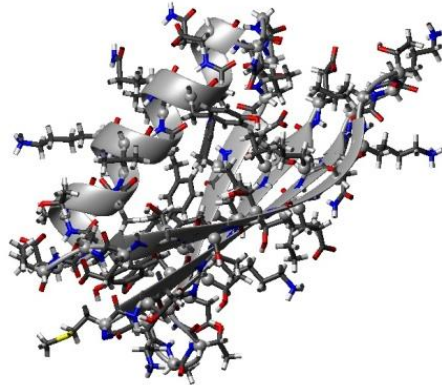
+

Experimental restraints: NOE, RDC, and etc.

+

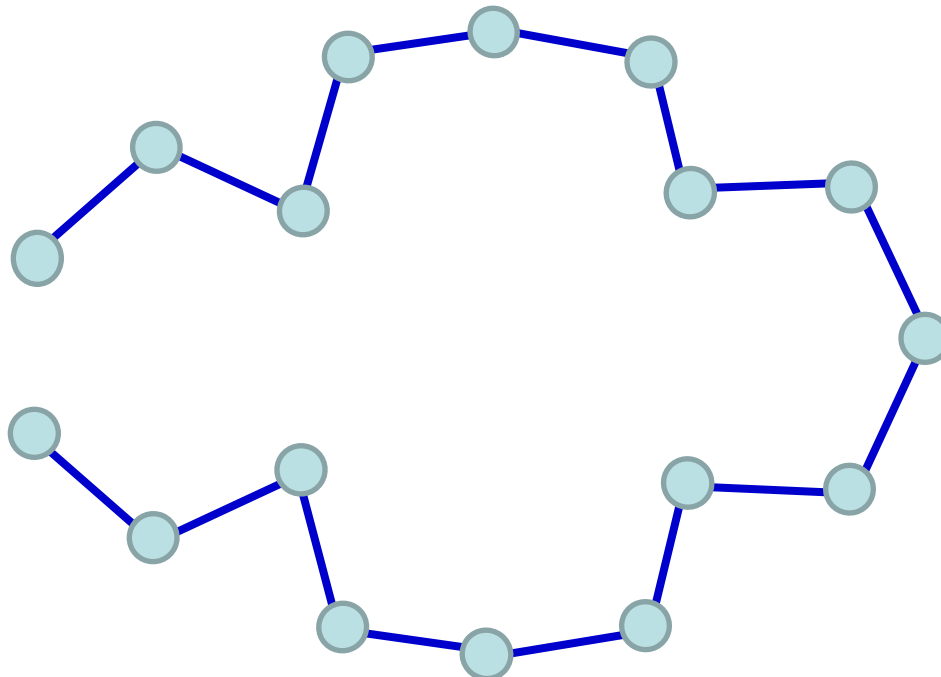
*a priori* restraints from database of known structures

=



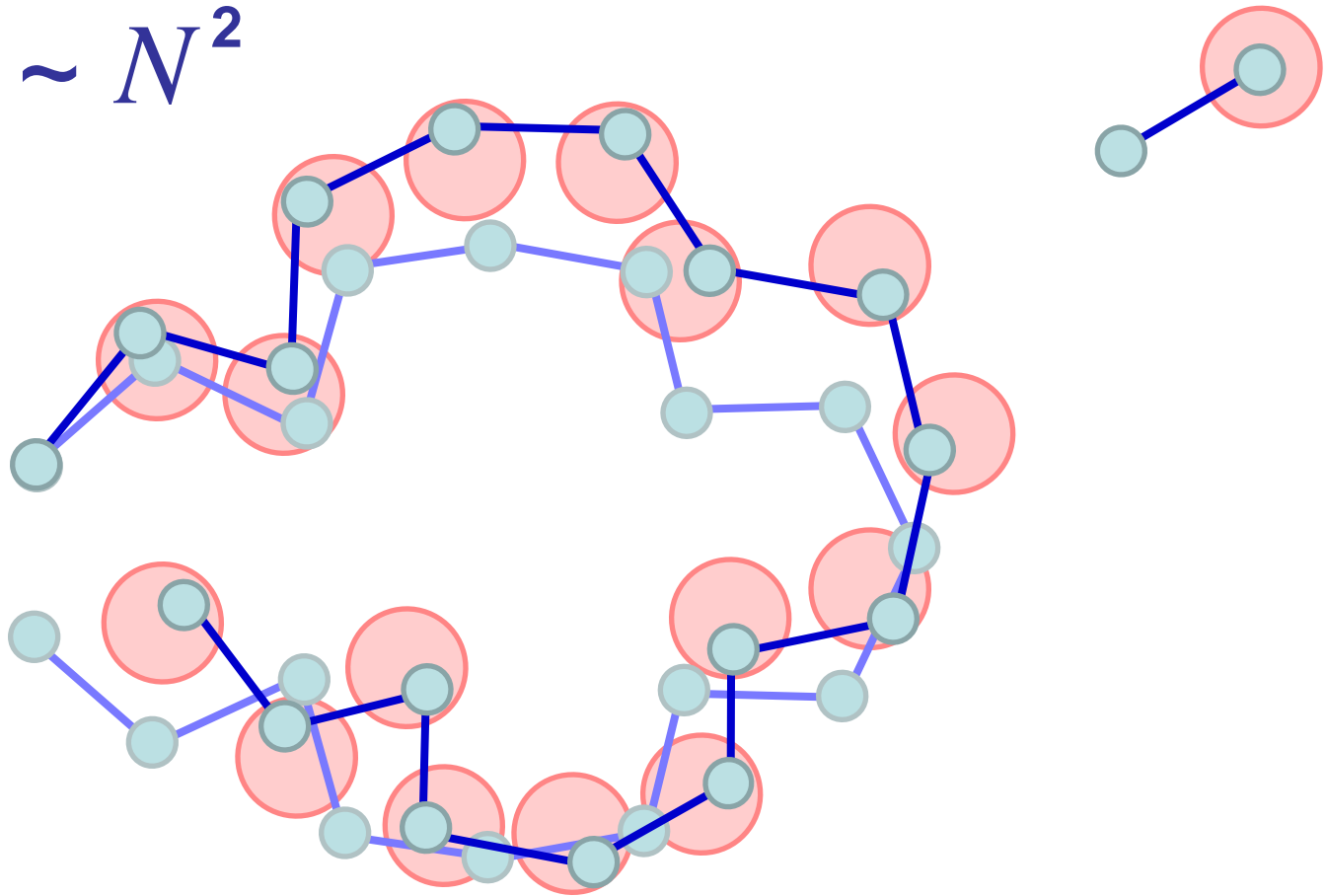
# Local and Global restraints

Ideal structure



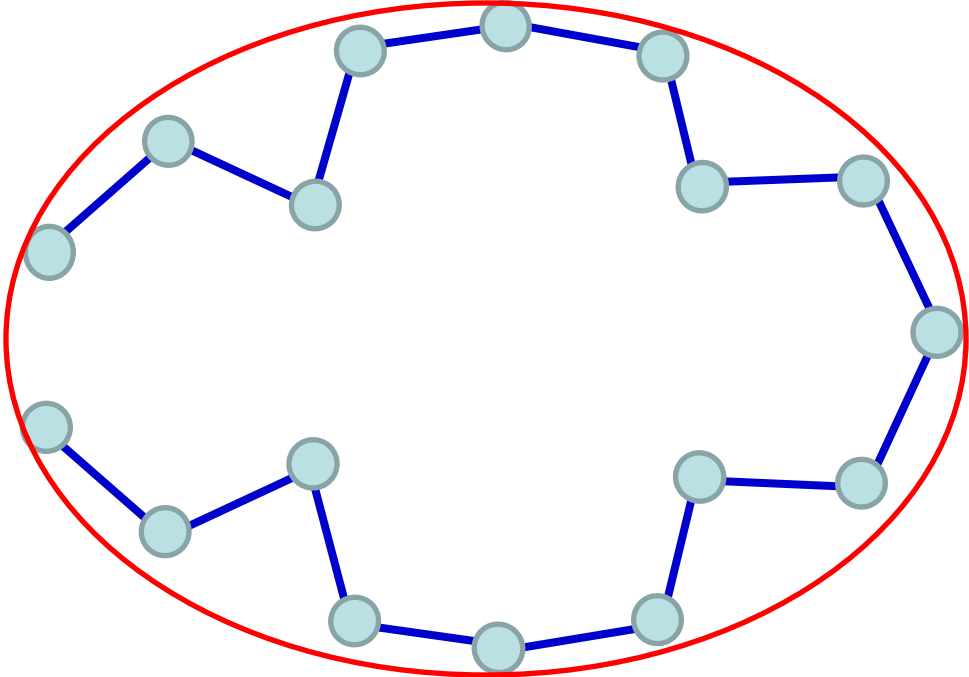
# Local restraints

$$\Delta \sim N^2$$



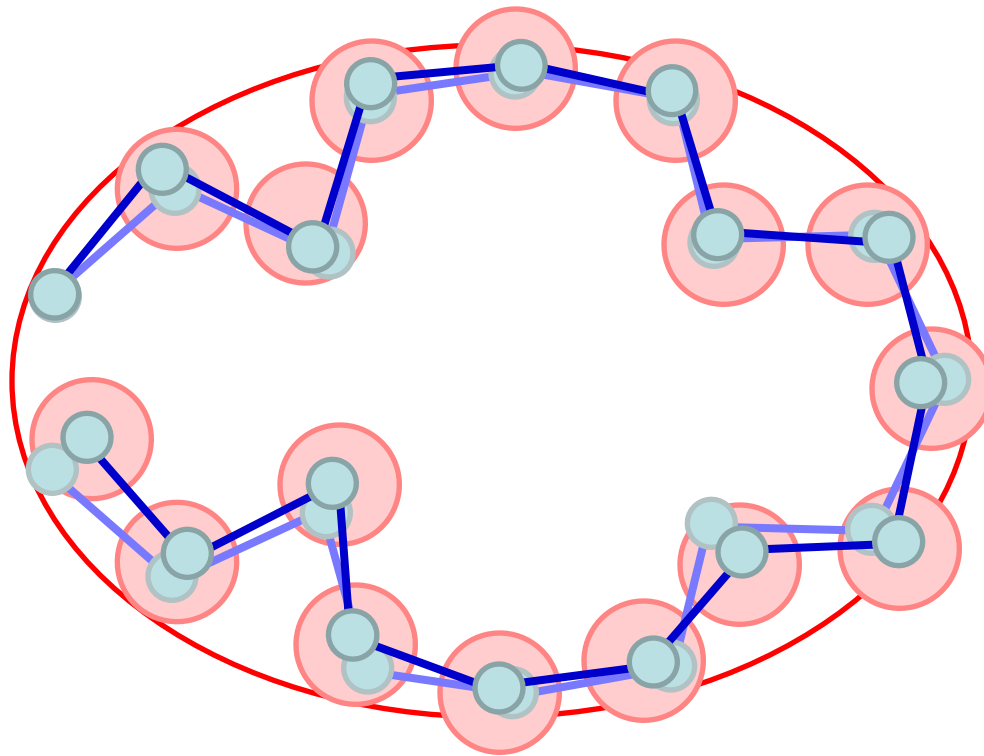
# Global restraints

# Overall shape





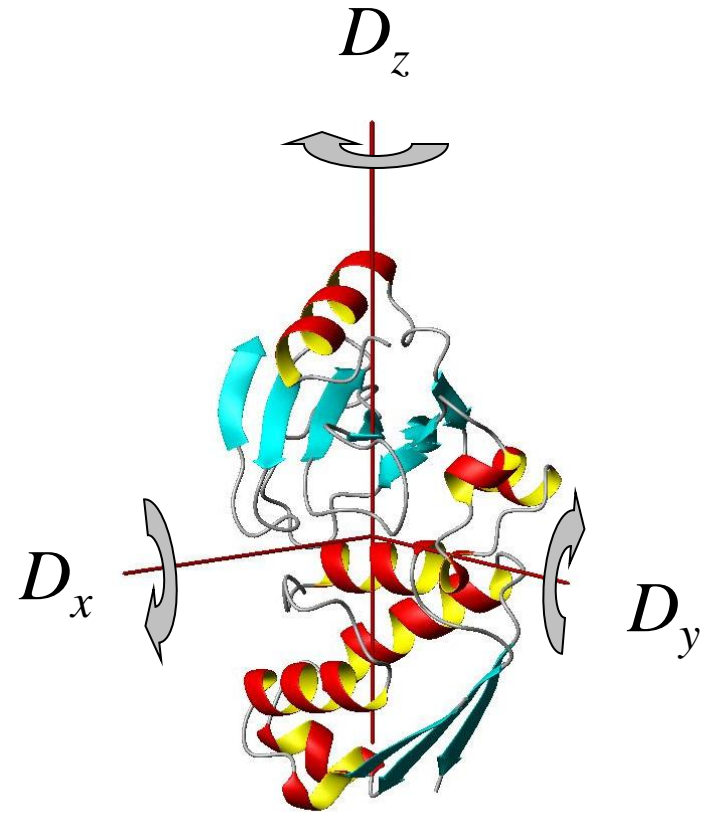
# Local and Global restraints



# Overall shape restraints from protein dynamics

$$\begin{bmatrix} D_x & 0 & 0 \\ 0 & D_y & 0 \\ 0 & 0 & D_z \end{bmatrix}$$

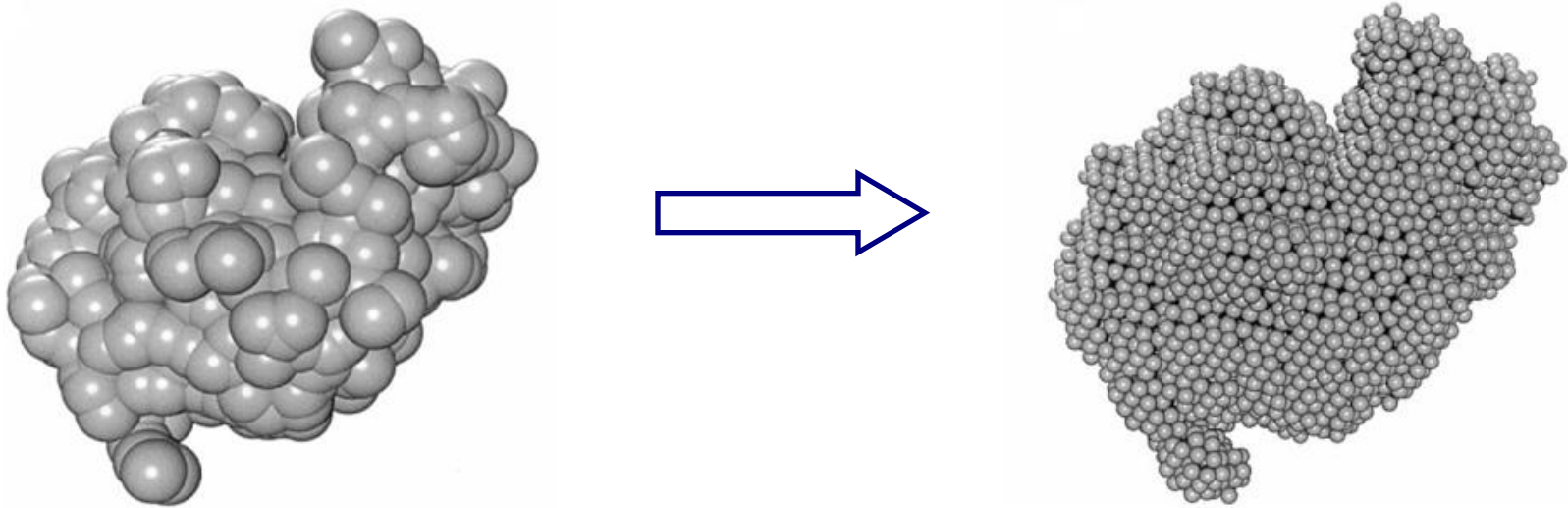
**3** Euler angles for  
**Diffusion Tensor PAF**



Using information about proteins shape encoded in protein rotation diffusion tensor for protein structure determination requires fast and accurate method for calculation protein rotation diffusion tensor for a given protein conformation

# Modeling **Diffusion Properties of Proteins**

## Bead algorithms



Extrapolation for Bead size  $\longrightarrow$  zero

**10 000 beads**

# Diffusion Properties of Proteins


## from ellipsoid model

Why an ellipsoid model ?

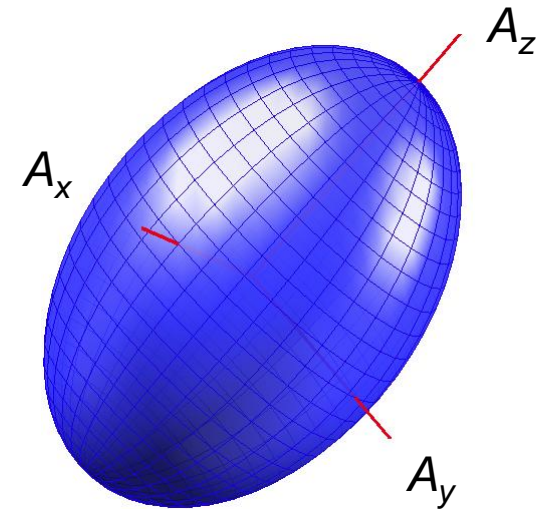
Diffusion Tensor

$$\begin{bmatrix} D_x & 0 & 0 \\ 0 & D_y & 0 \\ 0 & 0 & D_z \end{bmatrix}$$

One-to-One  
mapping



Ellipsoid Shell



**3 Euler angles for  
Diffusion Tensor PAF**

**3 Euler angles for  
Ellipsoid orientation**

# Diffusion Properties of Proteins

## from ellipsoid model

Main problem:

How to build the ellipsoid shell for a protein structure ?

State of the art: Inertia-equivalent ellipsoids

$$Re = \frac{\rho V L}{\mu} = \frac{\text{Inertia Forces}}{\text{Friction Forces}} \approx 0.01$$

$$\tau_{inertial\ relaxation} \approx 10^{-13} s$$

$$d_{inertial\ relaxation} \approx 0.1 \text{ \AA}$$

**Inertia is irrelevant** for protein diffusion

**Cantor & Schimmel** (1980)

# Diffusion Properties of Proteins

## from ellipsoid model

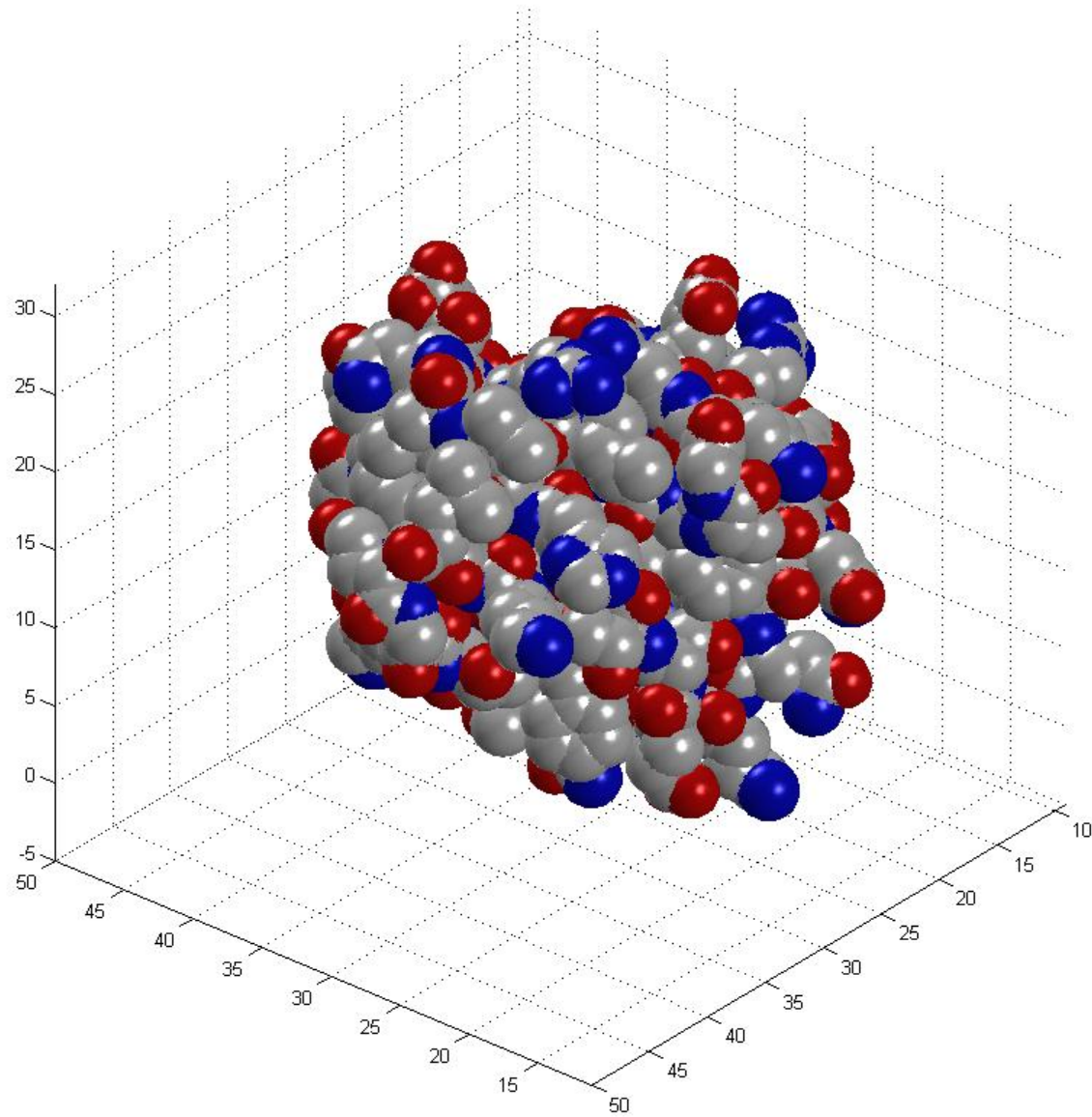
**Diffusion process** related to **friction**

The **friction** occurs at protein **surface**

## Proposal

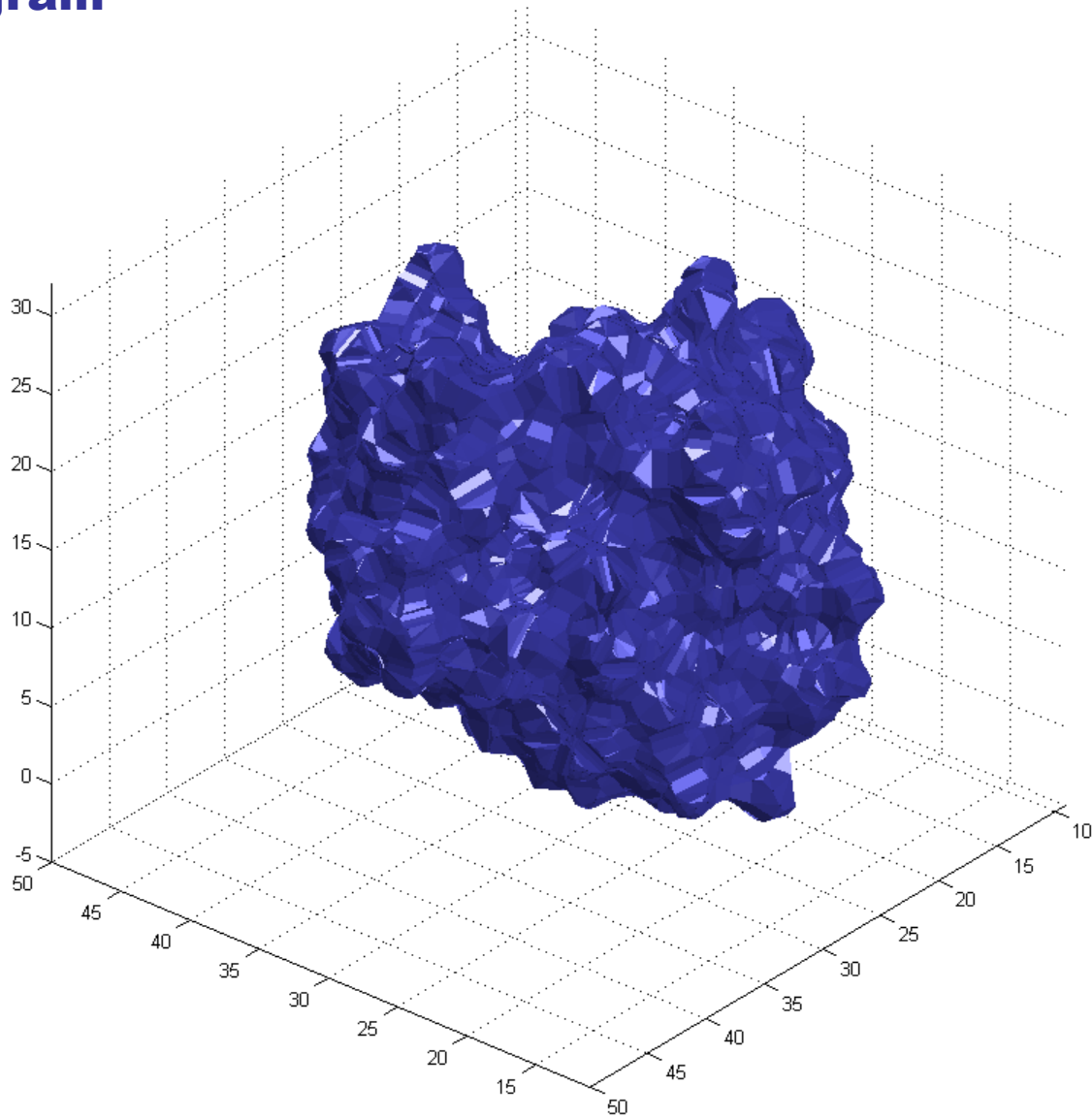
Let us use topology of protein surface to derive  
Equivalent ellipsoid for protein

## Mapping protein surfaces





# Mapping protein surfaces SURF program



# Build equivalent ellipsoid

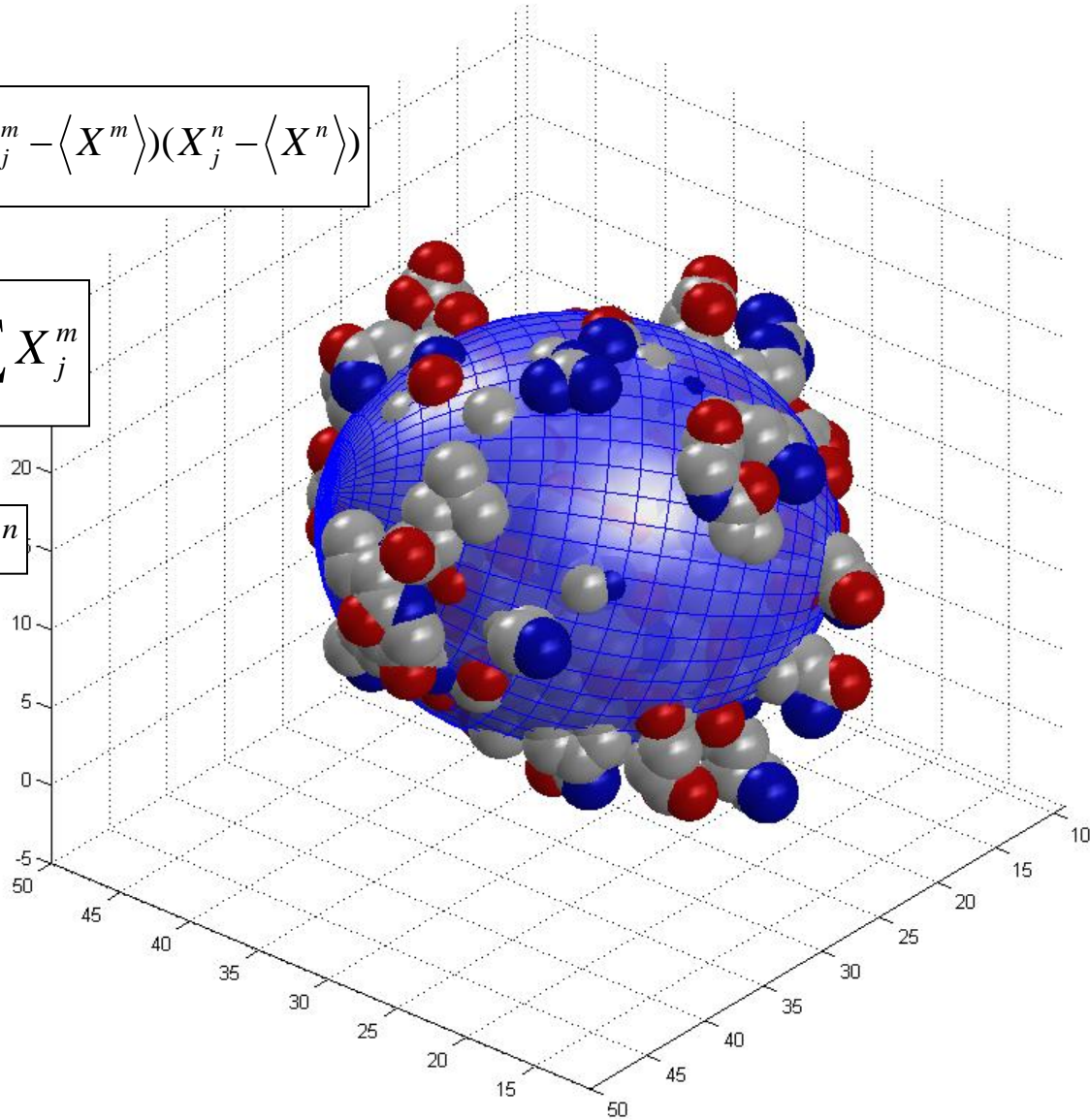
## Principal Component Analysis (PCA)

$$\text{Cov}_{m,n} = \frac{1}{N} \sum_{j=1}^N (X_j^m - \langle X^m \rangle)(X_j^n - \langle X^n \rangle)$$

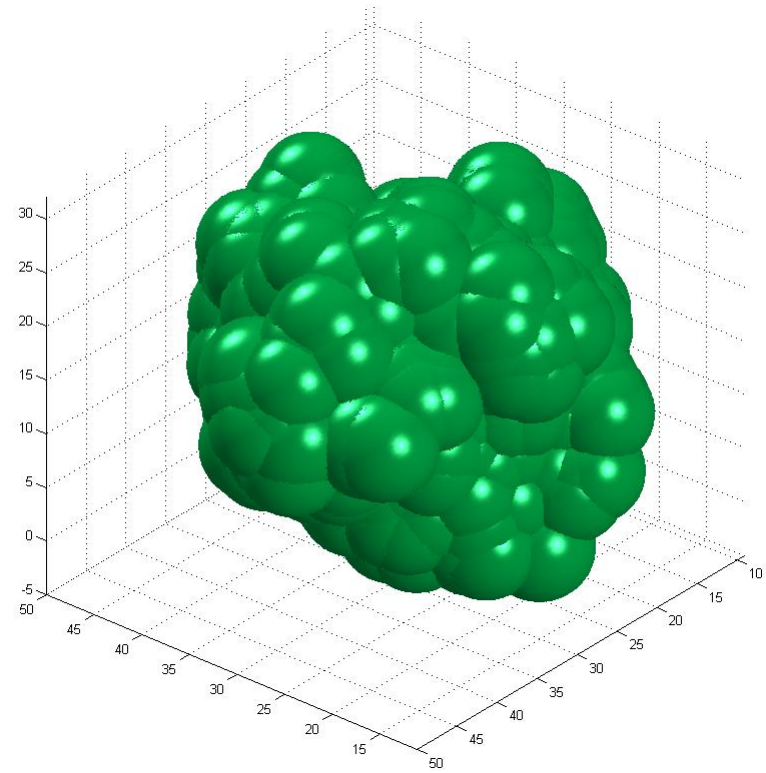
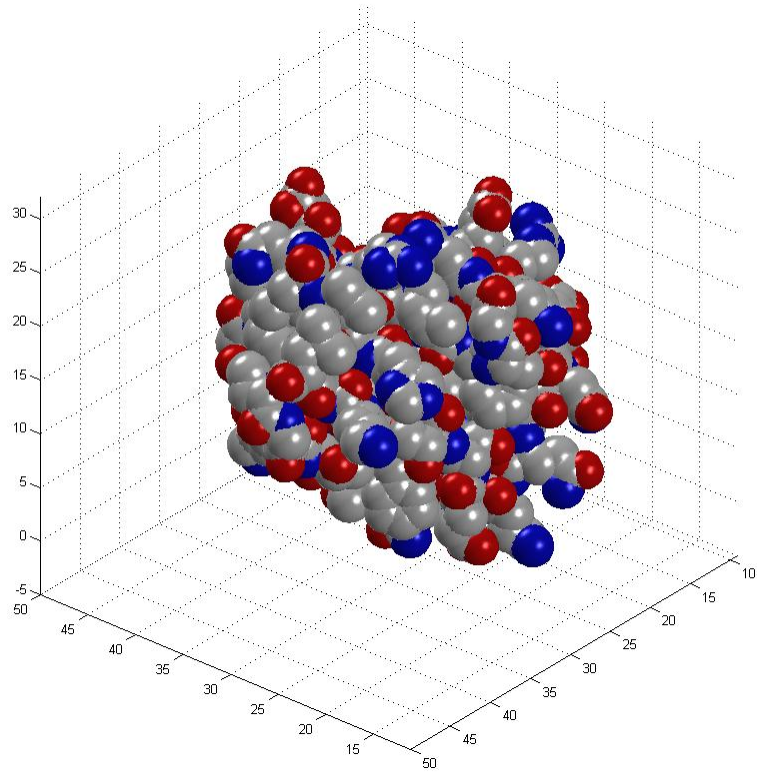
$$\langle X^m \rangle = \frac{1}{N} \sum_{j=1}^N X_j^m$$

$$\text{Cov} \mathbf{S}^n = \mathbf{E}^n \mathbf{S}^n$$

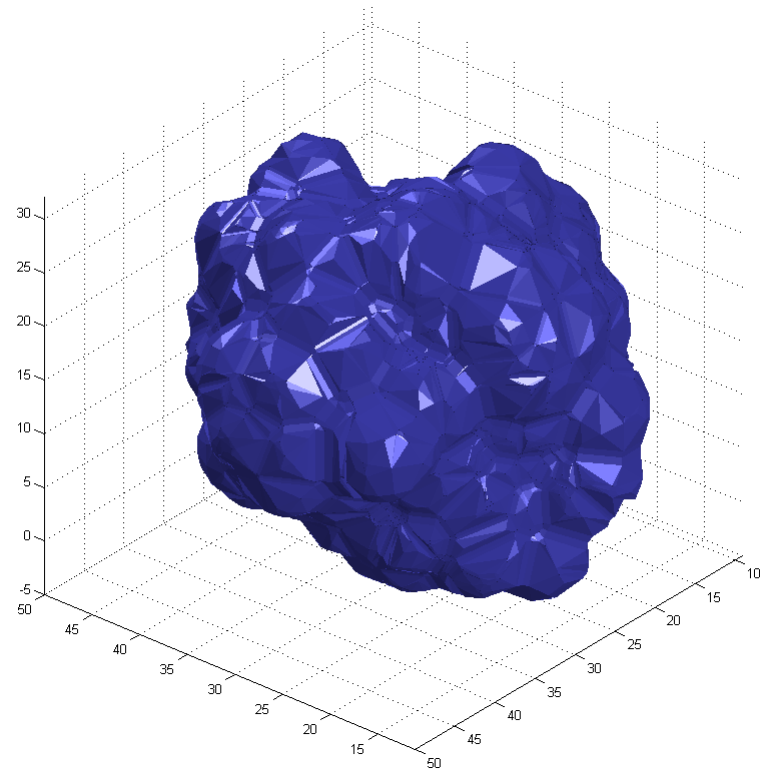
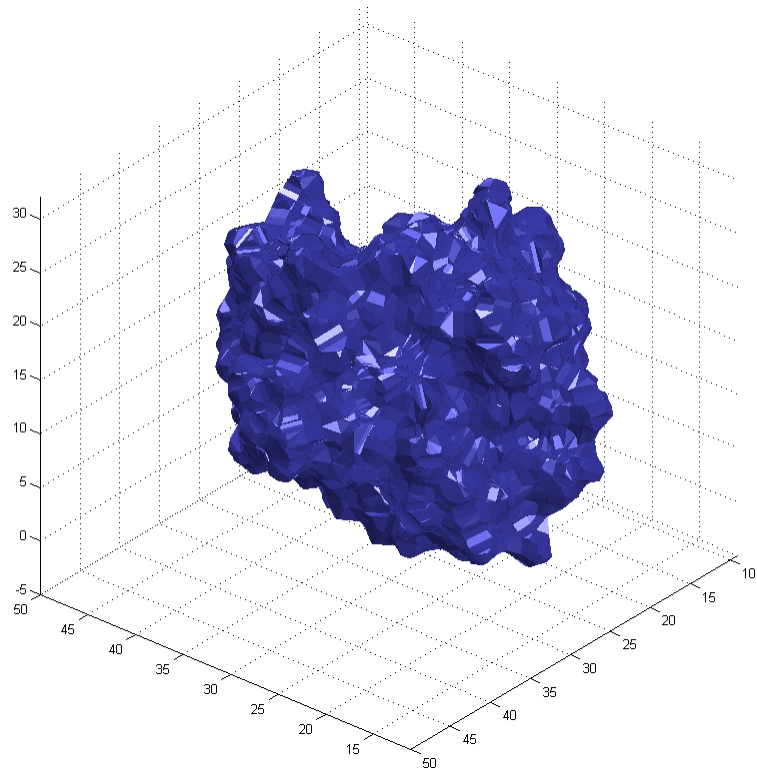
$$a_n = \sqrt{3E^n}$$



# Hydration shell

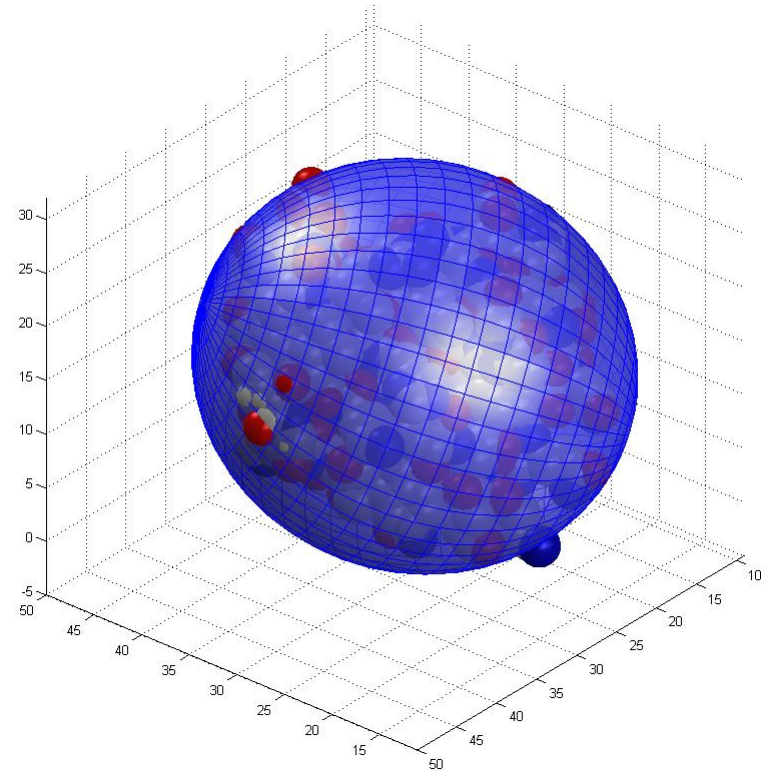
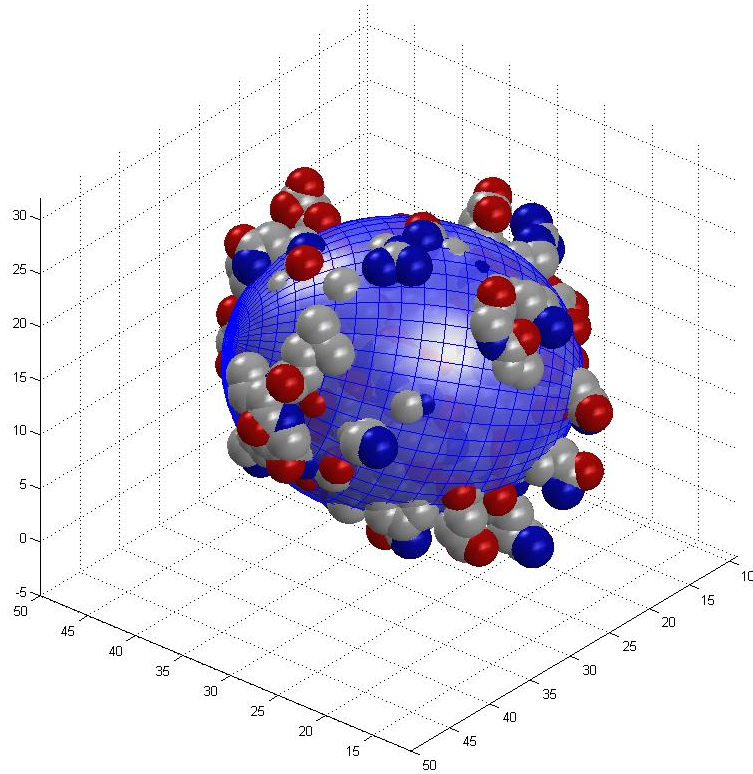


# Hydration shell



## Hydration shell

**Equivalent ellipsoid is approximately twice bigger**



# Complexity of the algorithms

ELM

$N_{at}$

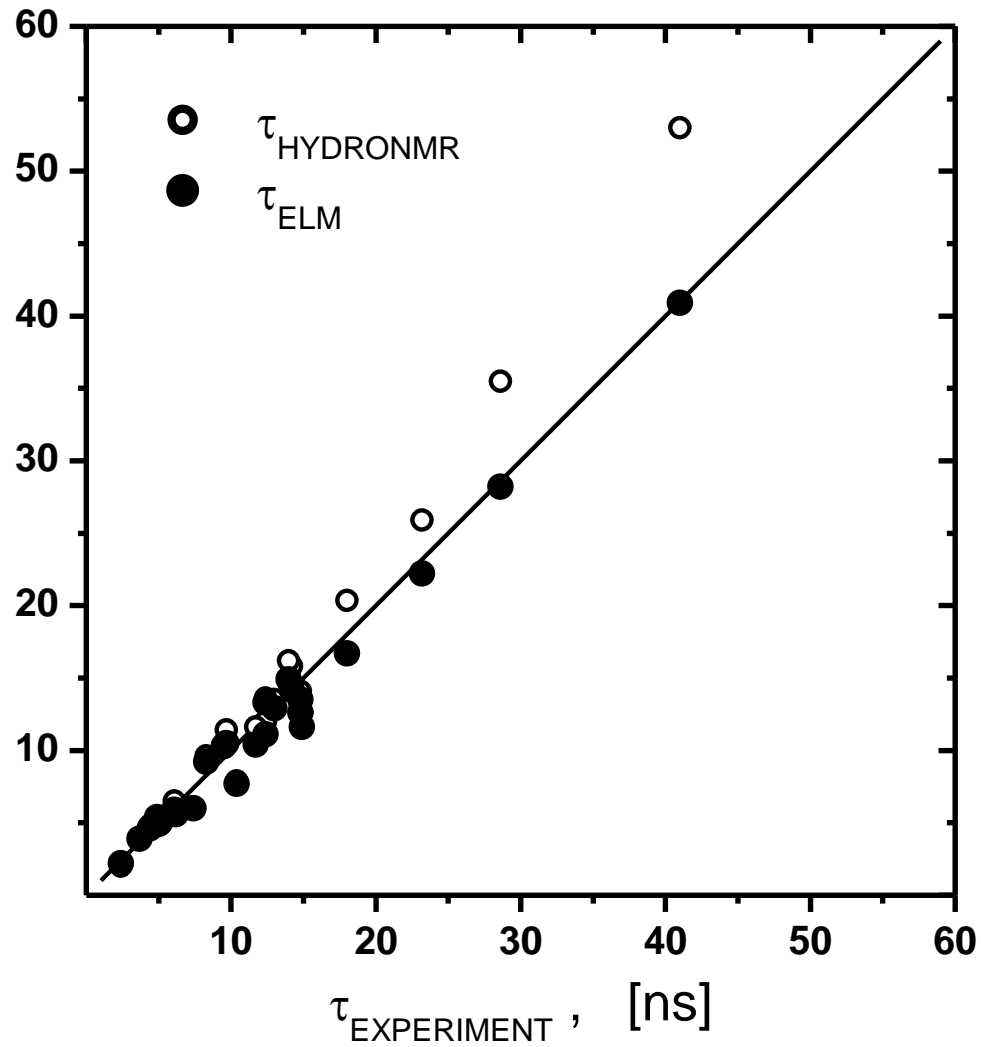
HYDRONMR

$N_{at}^2$

ELM : HYDRONMR

**1 : 500**

## Comparison with the experimental data



# ELM Algorithm

## Build equivalent ellipsoid with PCA

$$Cov_{m,n} = \frac{1}{N} \sum_{j=1}^N (X_j^m - \langle X^m \rangle)(X_j^n - \langle X^n \rangle) \quad \langle X^m \rangle = \frac{1}{N} \sum_{j=1}^N X_j^m$$

$$a_n = \sqrt{3E^n} \quad \mathbf{CovS}^n = \mathbf{E}^n \mathbf{S}^n$$

## Evaluate diffusion tensor components with Perrin's Equations

$$D_l = \frac{kT}{C_l} \quad C_x = \frac{16\pi\eta(a_y^2 + a_z^2)}{3(a_y^2 Q + a_z^2 R)} \quad C_y = \frac{16\pi\eta(a_x^2 + a_z^2)}{3(a_z^2 R + a_x^2 P)} \quad C_z = \frac{16\pi\eta(a_x^2 + a_y^2)}{3(a_x^2 P + a_y^2 Q)}$$

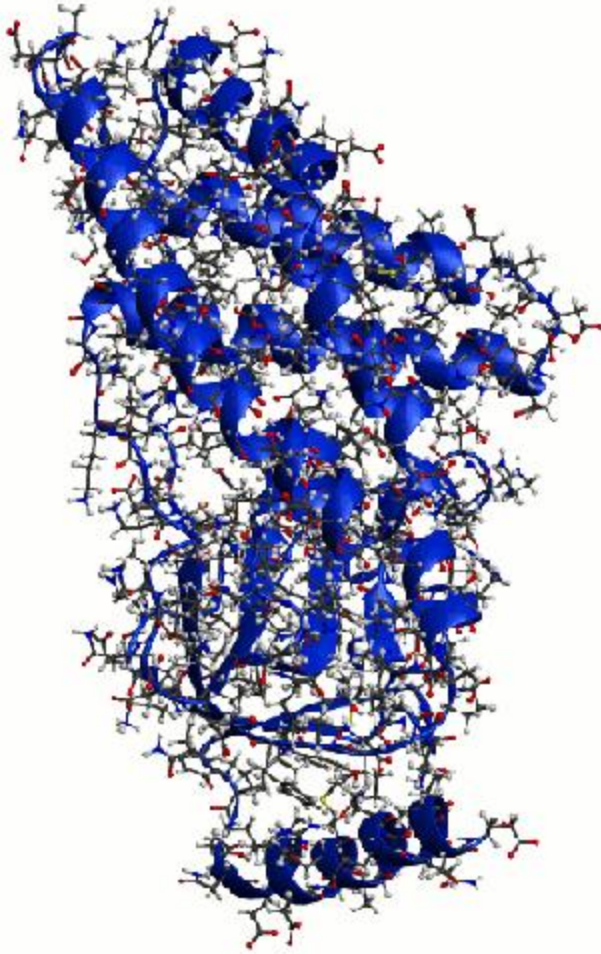
$$P = \int_0^\infty \frac{ds}{\sqrt{(a_x^2 + s)^3 (a_y^2 + s)(a_z^2 + s)}} \quad Q = \int_0^\infty \frac{ds}{\sqrt{(a_y^2 + s)^3 (a_z^2 + s)(a_x^2 + s)}} \quad R = \int_0^\infty \frac{ds}{\sqrt{(a_z^2 + s)^3 (a_x^2 + s)(a_y^2 + s)}}$$

Perrin J. *Phys. Radium* (1934, 1936)



Fast and accurate ELM algorithm, which is able to provide closed-form derivatives of the energy function associated with the overall shape restraints, can be used in Molecular Dynamics and for gradient minimization in protein structure determination routines.

# Refinement of a protein structure with Xplor-NIH using overall shape restraints from diffusion tensor



N terminal domain from Enzyme I (EIN)

Standard Xplor-NIH simulated annealing protocol started from 3000K down to 25 K  
With 12.5 K steps

Experimental restraints:

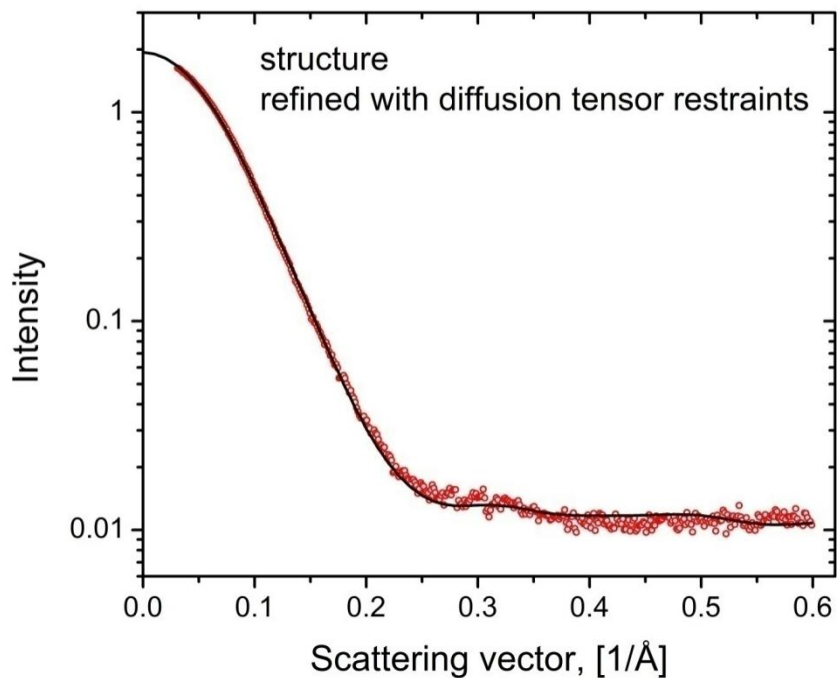
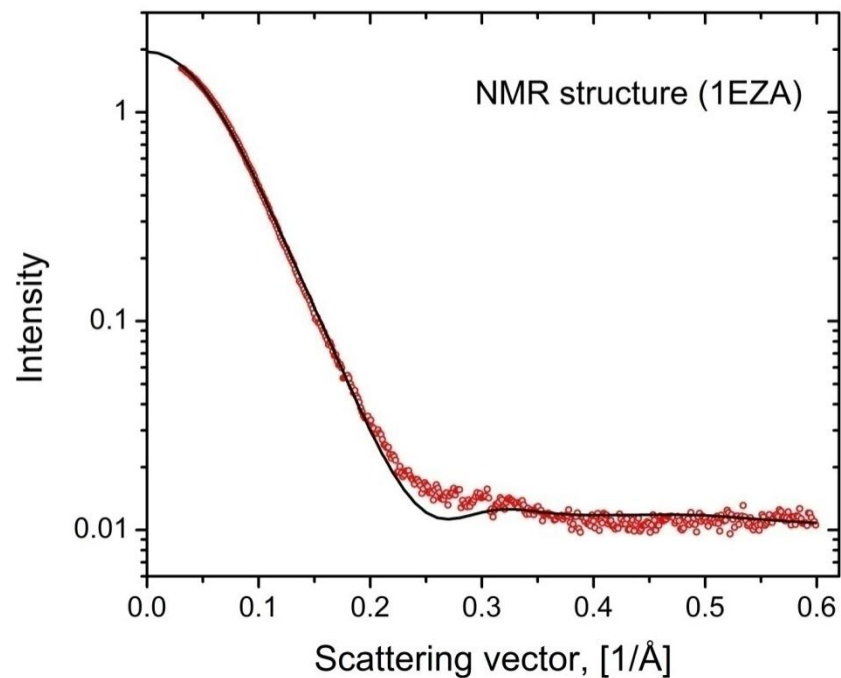
distance restraints derived from NOE

and

Components of Rotation Diffusion Tensor

10 lowest energy structures: Blue with diffusion tensor restraints  
Green without diffusion tensor restraints

## Validation with SAXS data



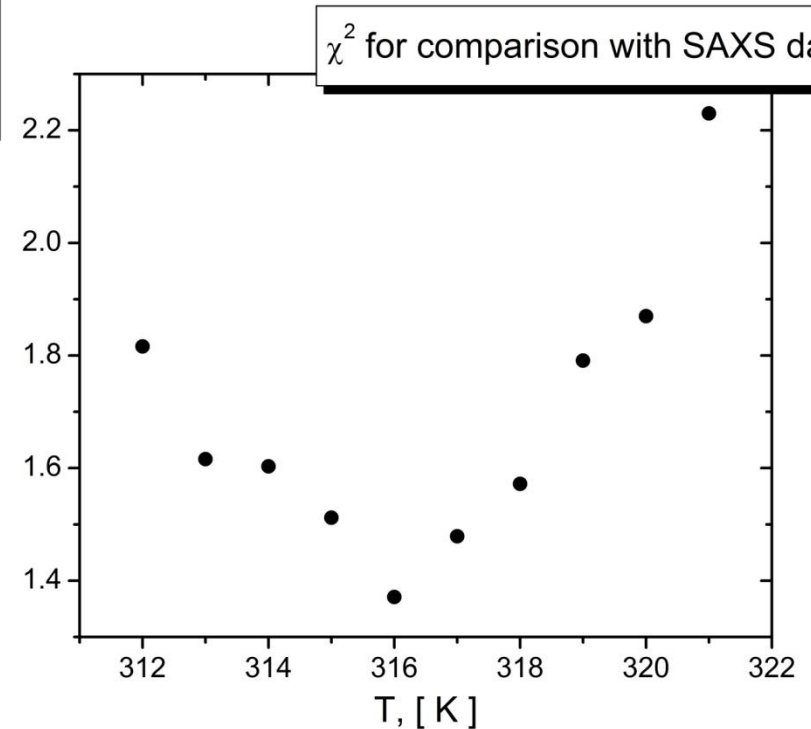
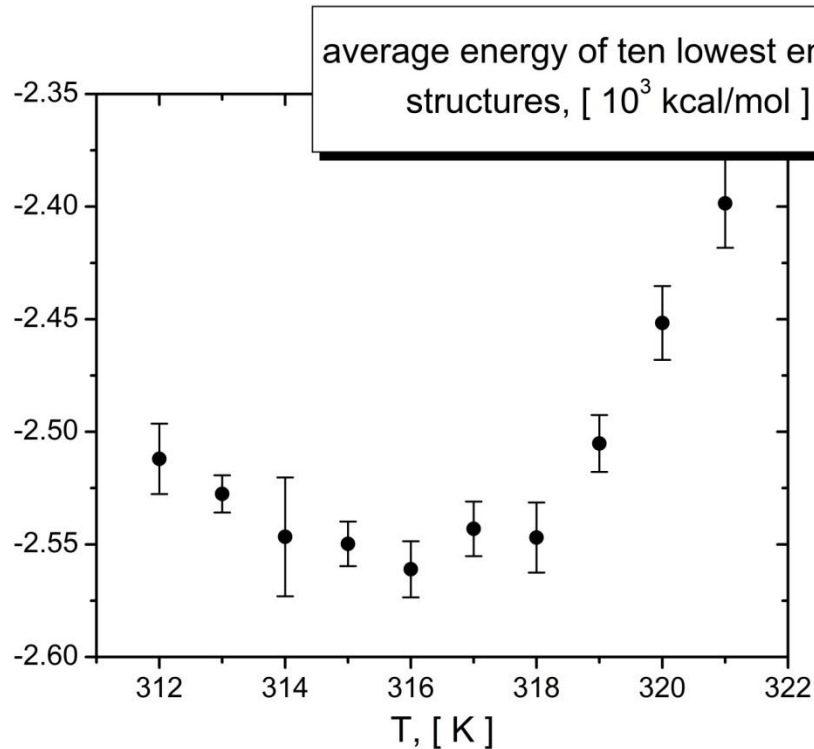
$\chi^2$  for comparison with SAXS data

1EZA (NMR)	2.03
1ZYM (X-ray)	1.38
Refined without diffusion tensor restraints	1.58
Refined with diffusion tensor restraints	1.37

## Effect of temperature settings for diffusion tensor term

Experimental temperature: 313 K

Temperature of the minimum: 316 K

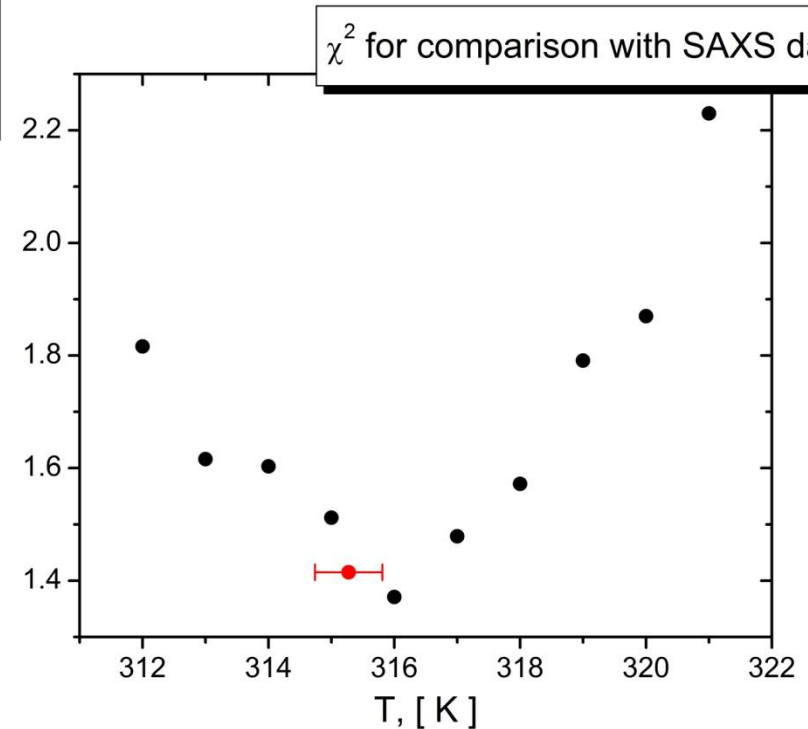
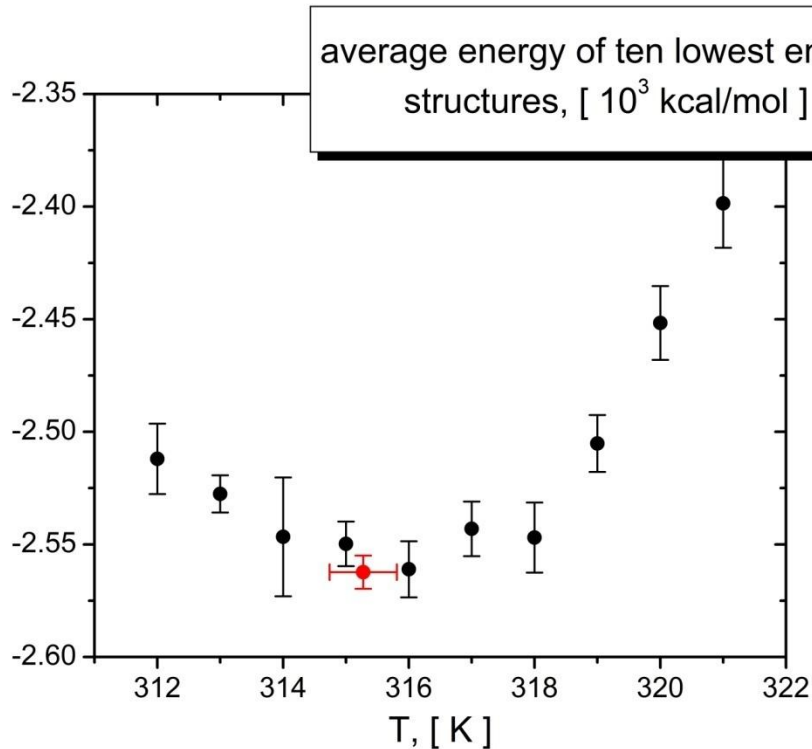


Uncertainties in  
thickness of hydration layer, sample temperature, and sample viscosity

# Effect of temperature settings for diffusion tensor term

Experimental temperature: 313 K  
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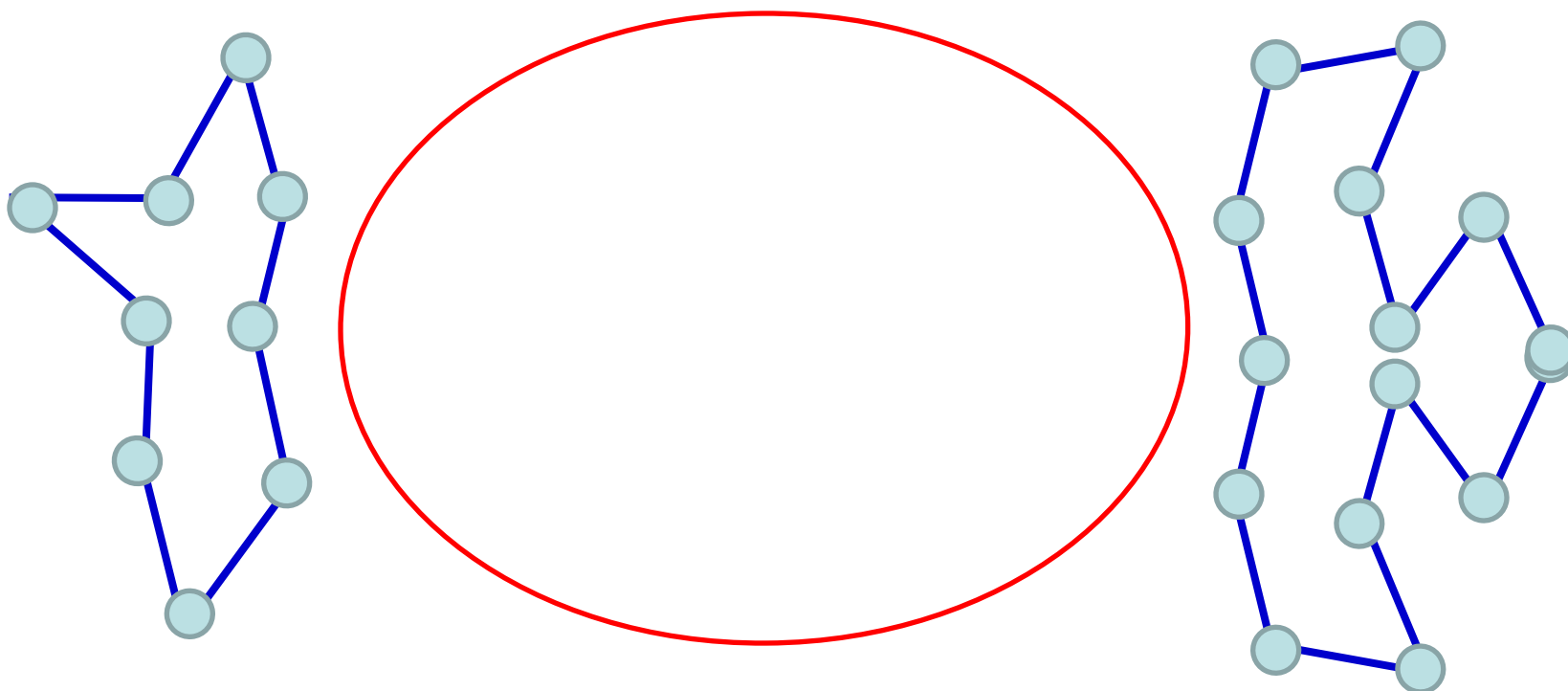
Optimized Temperature:  $315.3 \pm 0.5$



Uncertainties in  
thickness of hydration layer, sample temperature, and sample viscosity

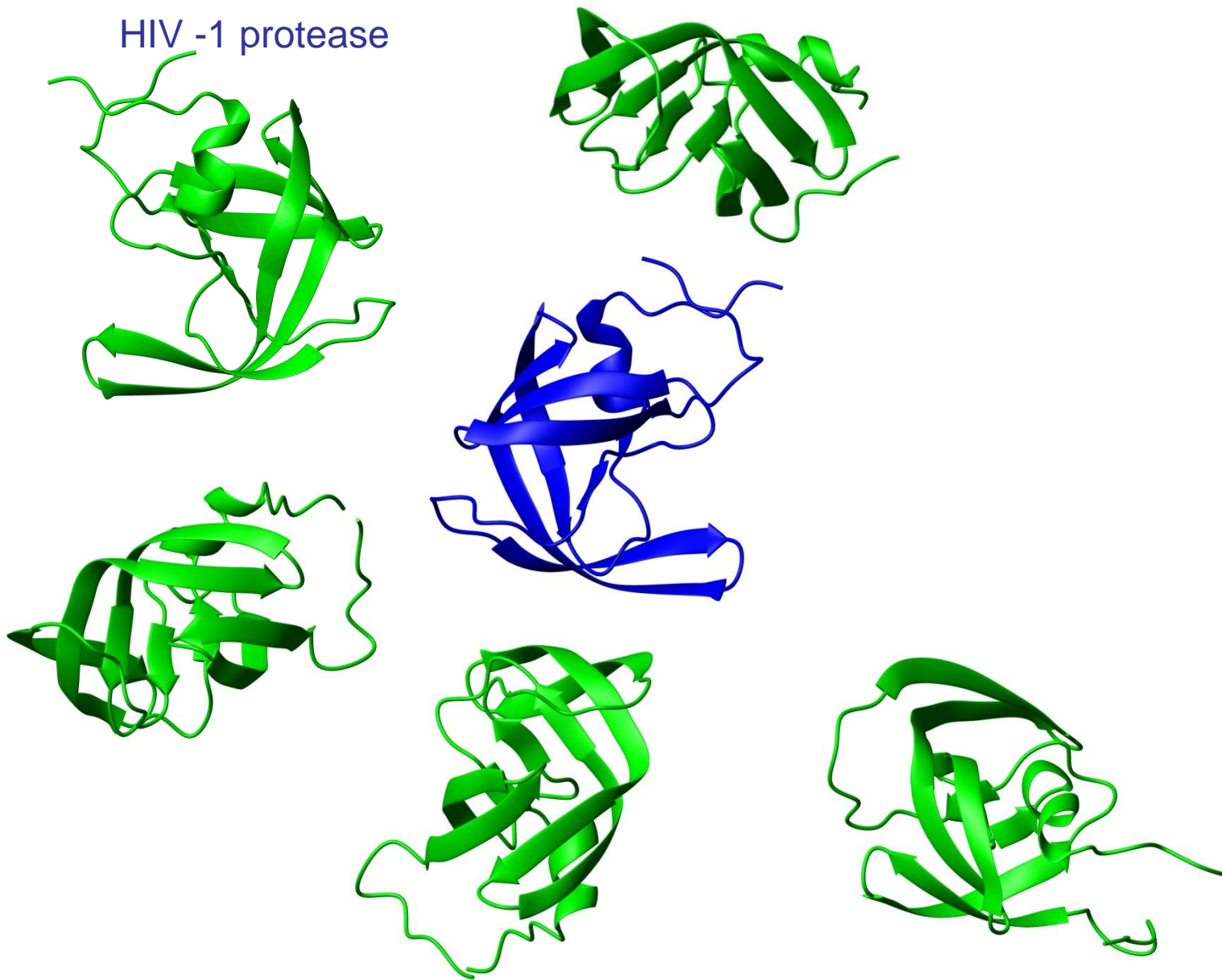
# Assembling structures of multi domain proteins using the overall shape restraints provided by the diffusion tensor

Global restraints on Overall shape

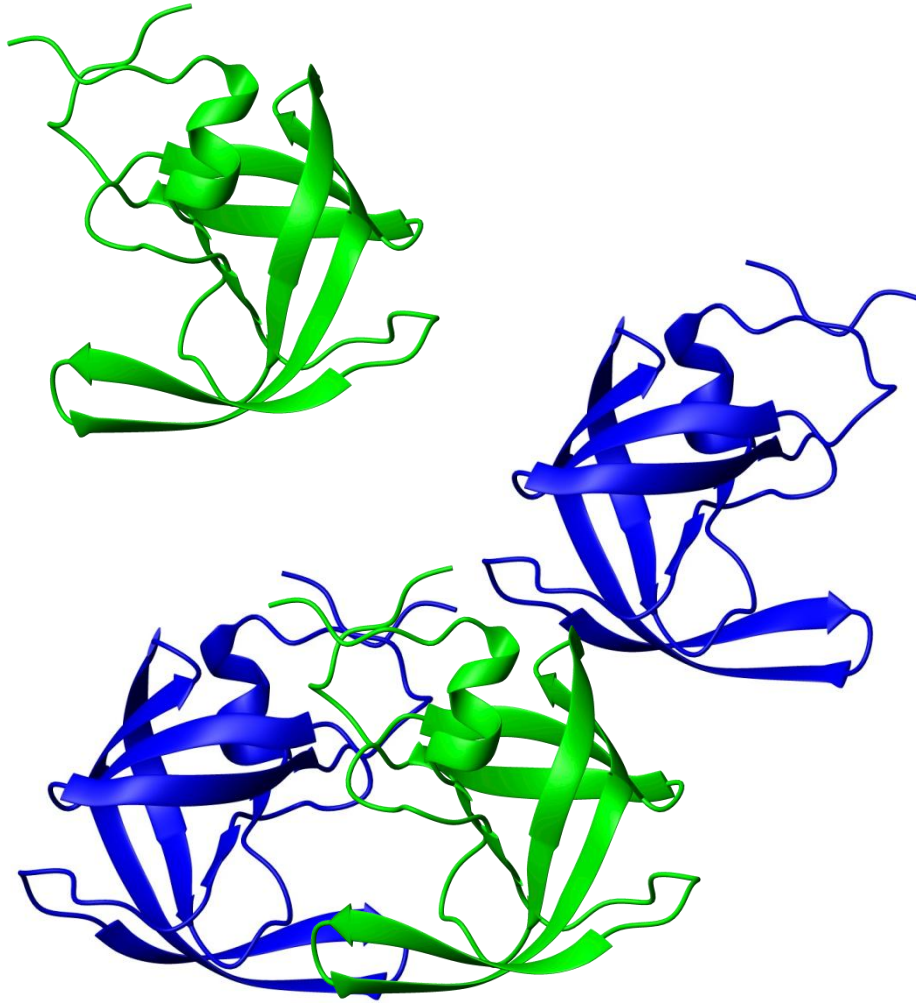


## Assembling structure of a symmetric protein homo dimer

HIV -1 protease



## Assembling structure of a symmetric protein homo dimer



Generic docking protocol

Part I:

Rigid body dynamics  
for raw domain positioning.

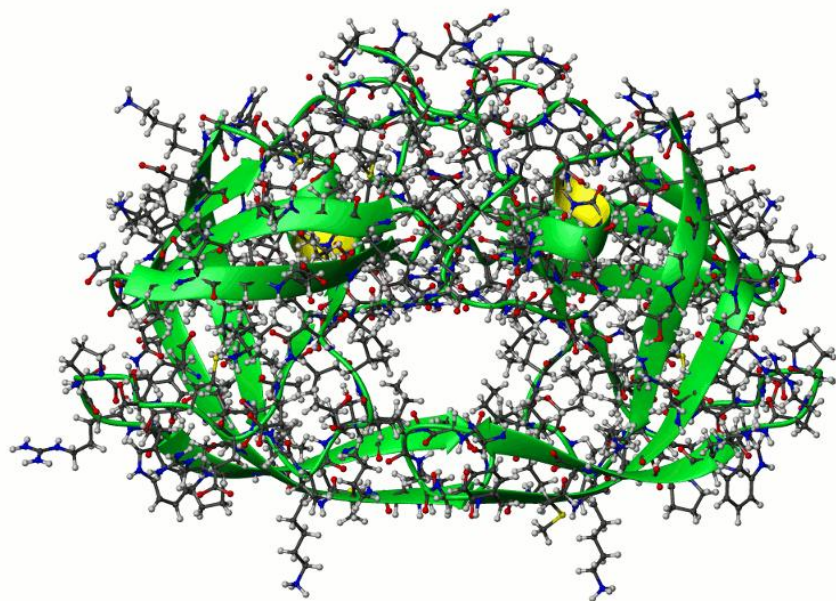
Part II:

Simulated annealing  
with flexible side chains  
for final adjustment.



# Assembling structure of a symmetric protein homo dimer

HIV -1 protease



10 lowest energy structures

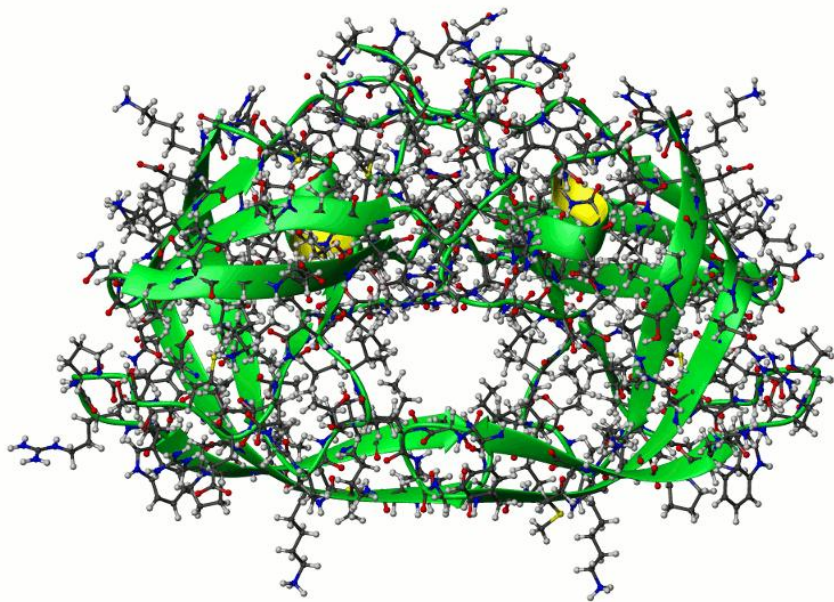
Randomization of domain positions and Rigid body dynamics repeated 10 times; then the lowest energy structure submitted to final simulated annealing part of the protocol

512 structures calculated.

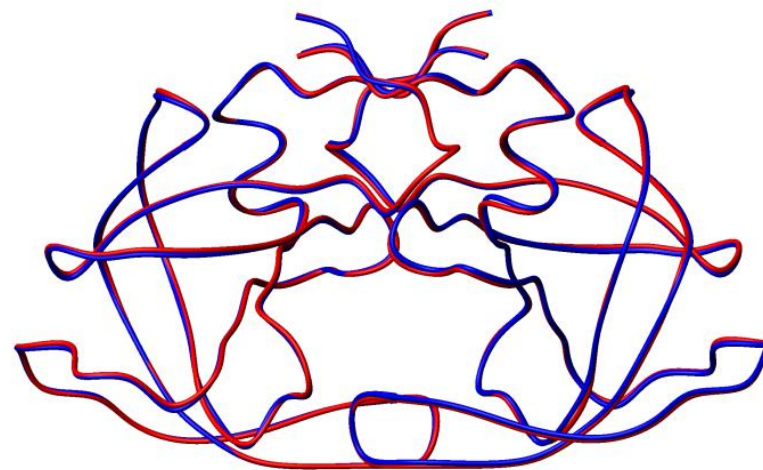
The only experimental restrains are Components of Rotation Diffusion Tensor

# Assembling structure of a symmetric protein homo dimer

HIV -1 protease



10 lowest energy structures



Averaged over 10 lowest energy structures (blue) versus reference (red)

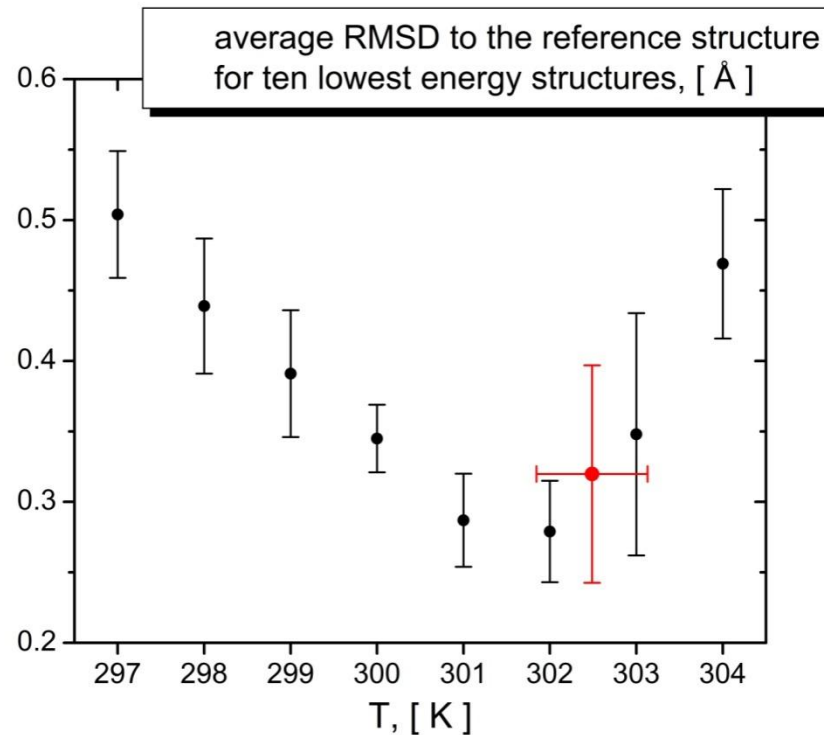
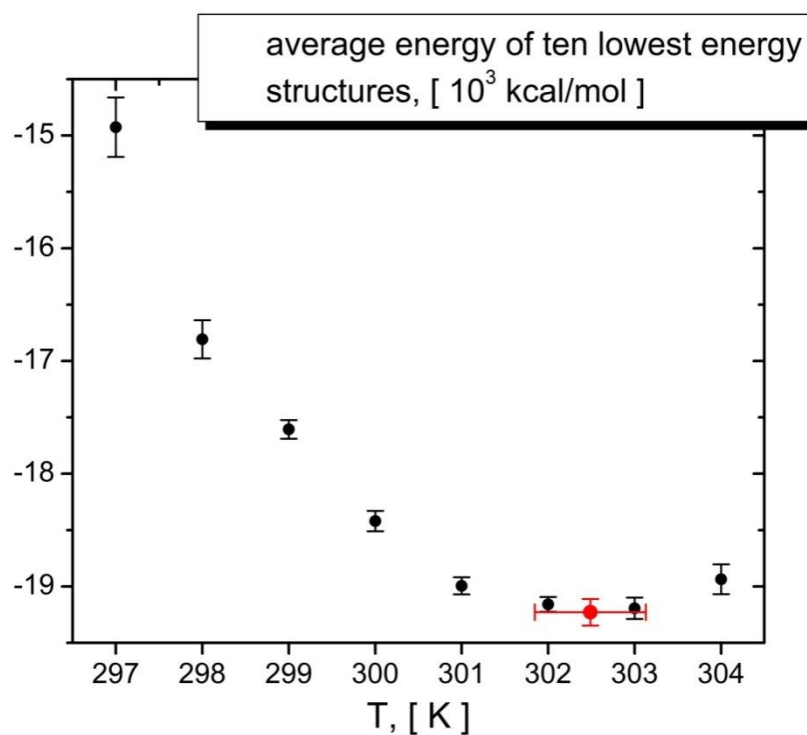
$C_{\alpha}$  RMSD  $0.35 \pm 0.09$  [Å]

## Assembling structure of a symmetric protein homo dimer

Experimental temperature: 300 K

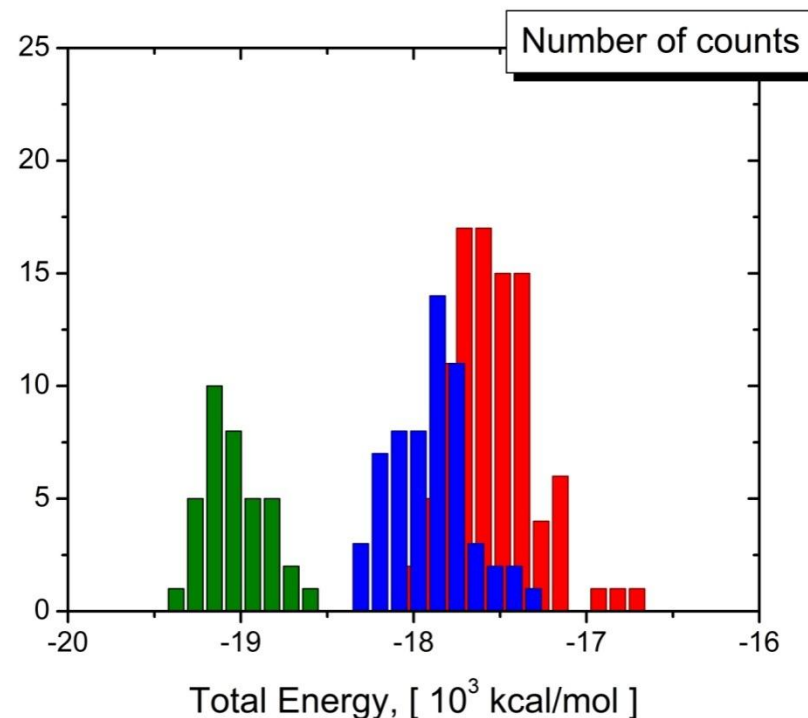
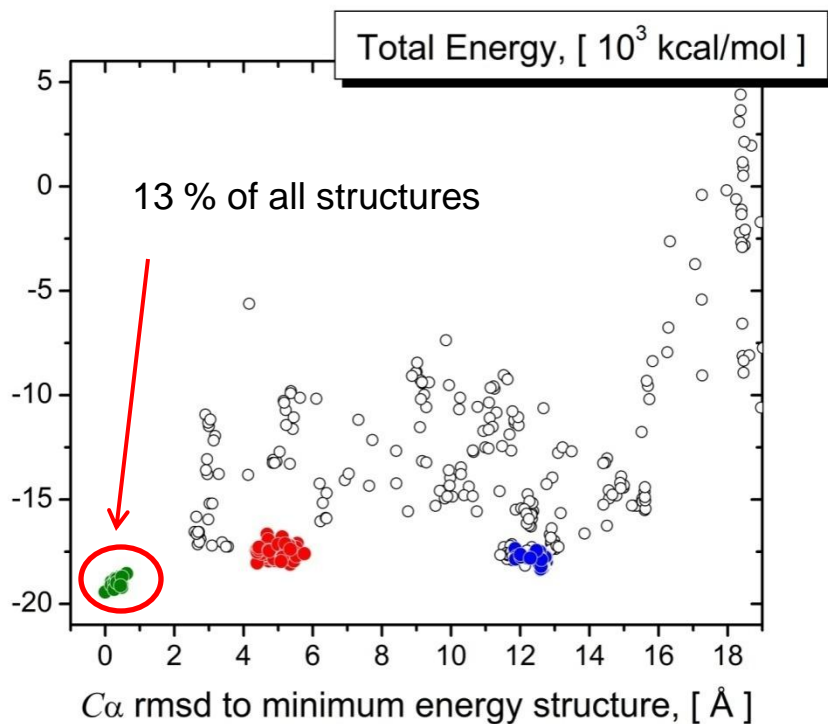
Temperature of the minimum: 303 K

Optimized Temperature:  $302.5 \pm 0.6$



# Assembling structure of a symmetric protein homo dimer

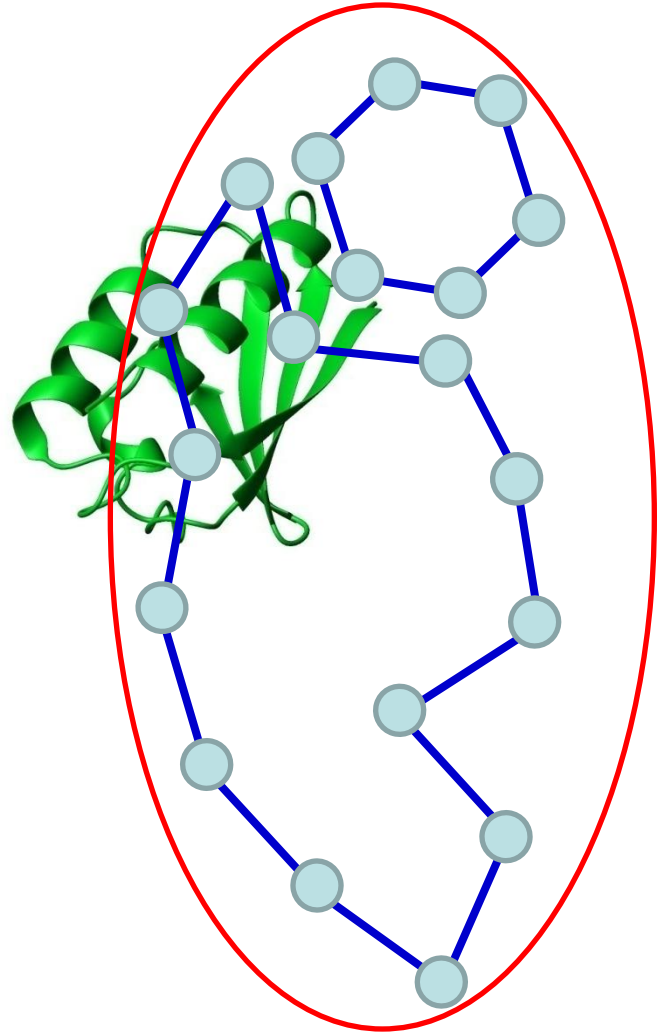
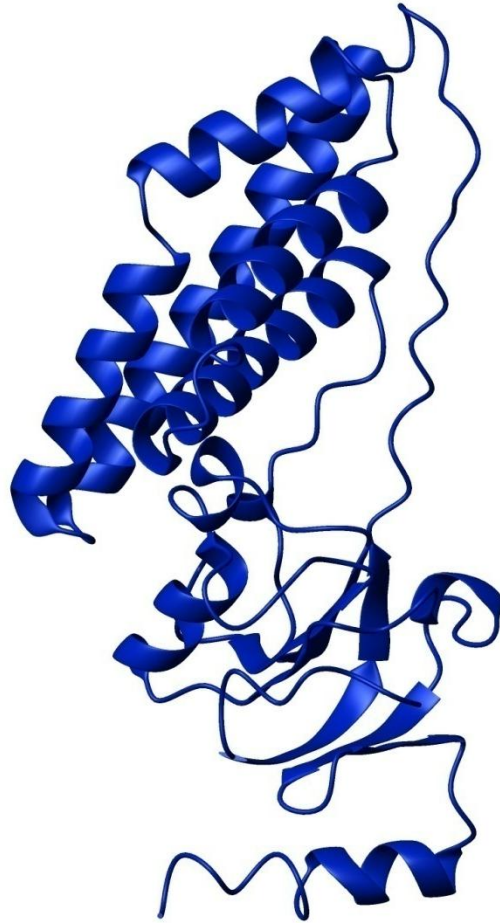
HIV -1 protease



Diffusion tensor shape restraints have they own right to define domain assembly in protein complexes

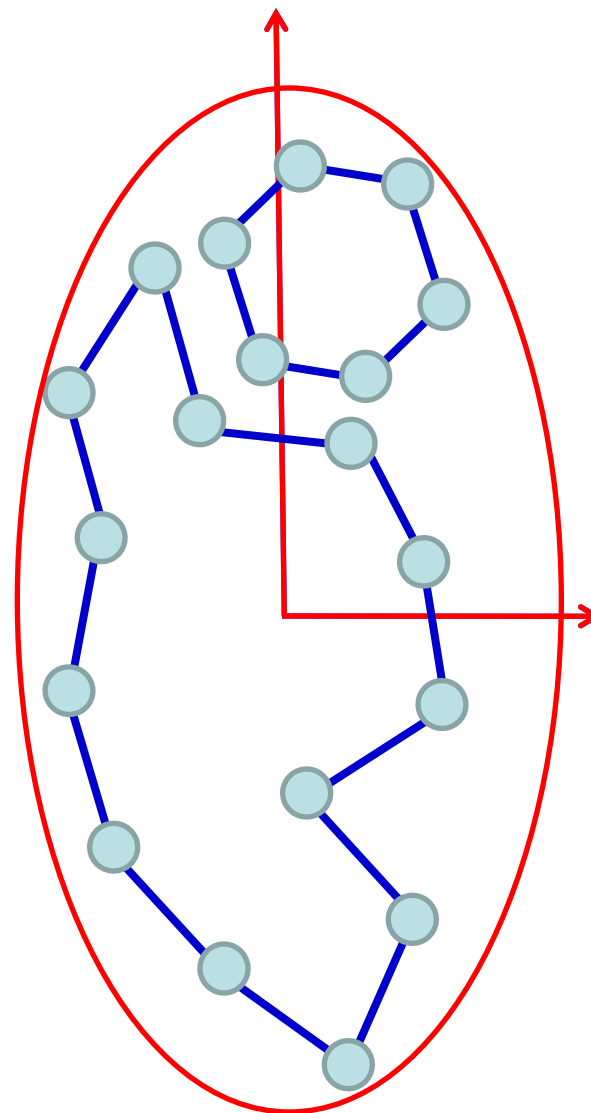
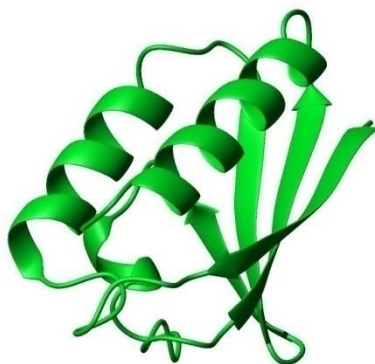
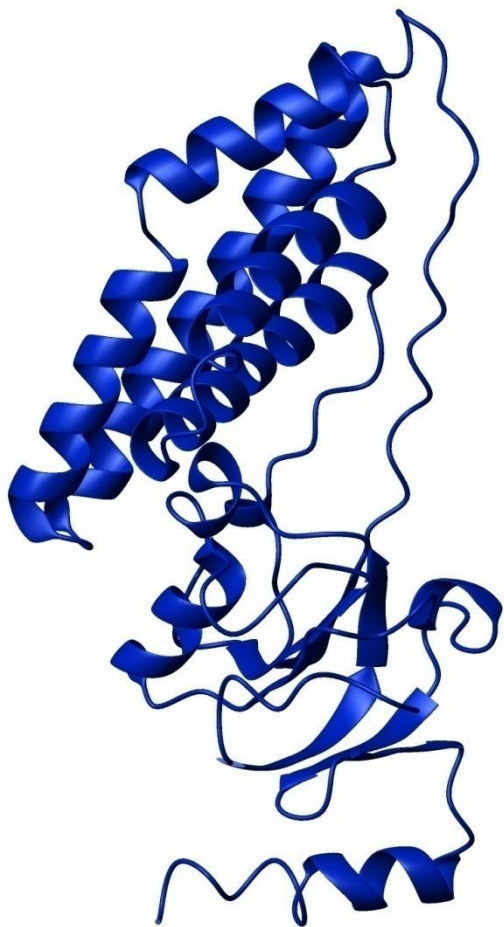
## Application to an asymmetric complex

EIN – HPr complex



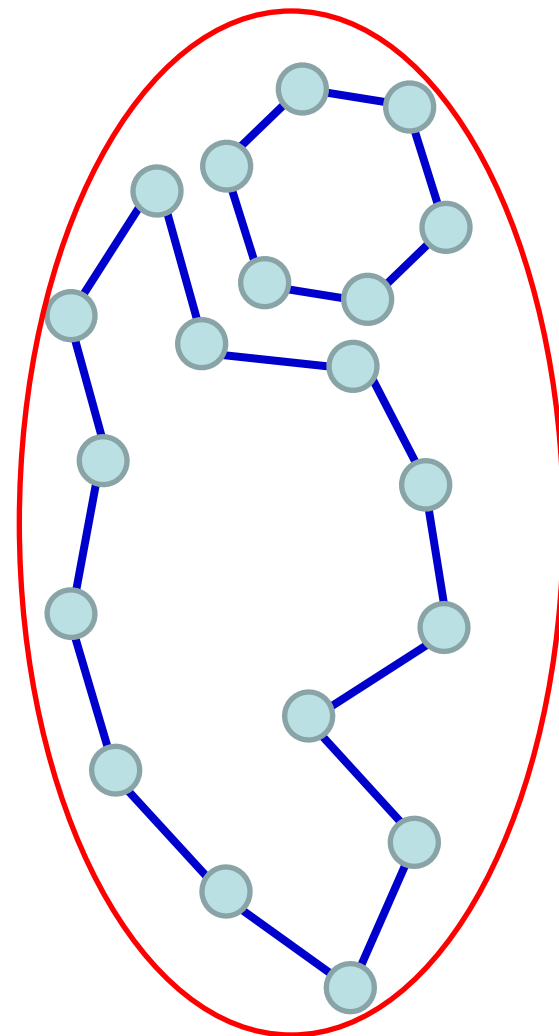
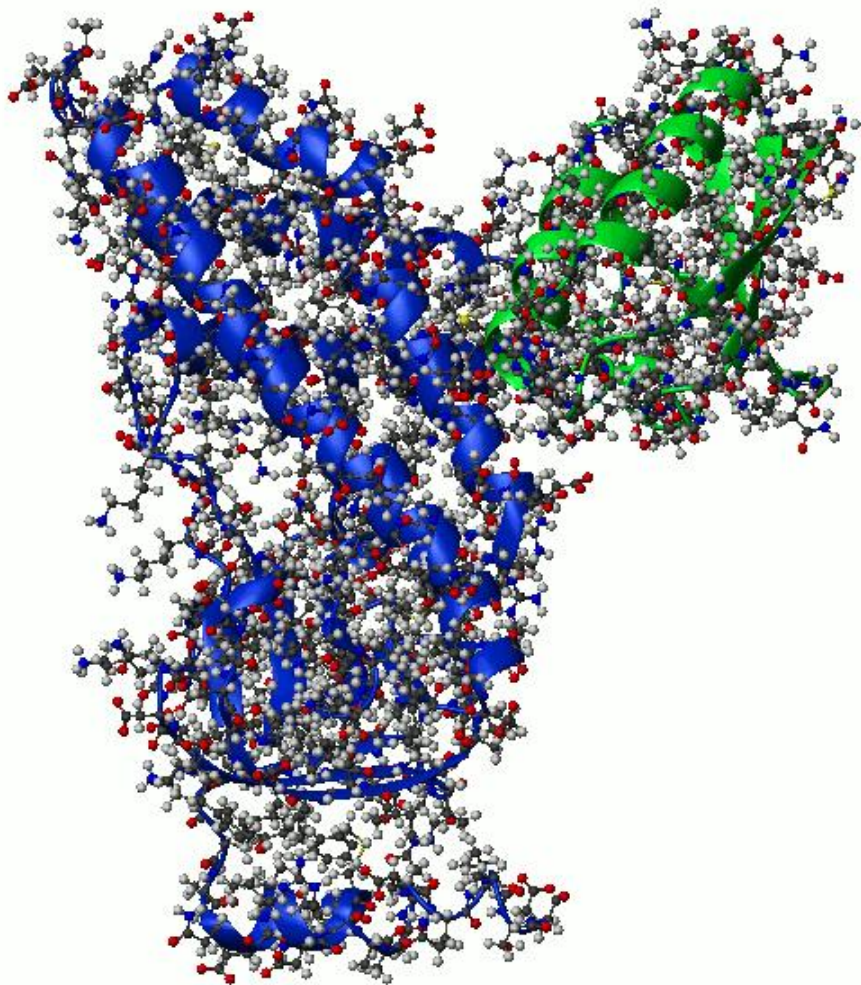
## Application to an asymmetric complex

EIN – HPr complex



## Application to an asymmetric complex

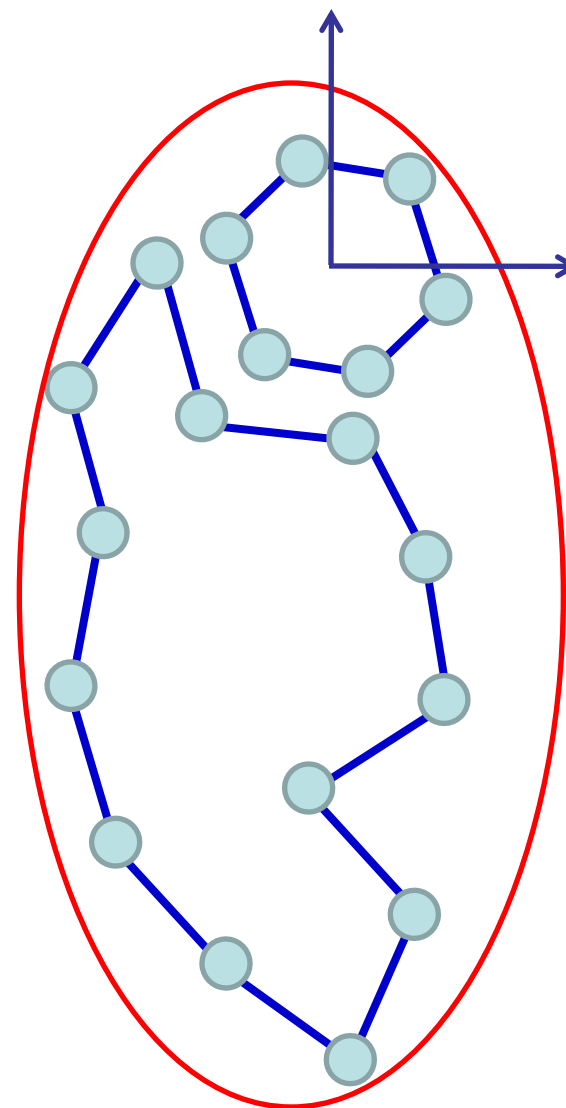
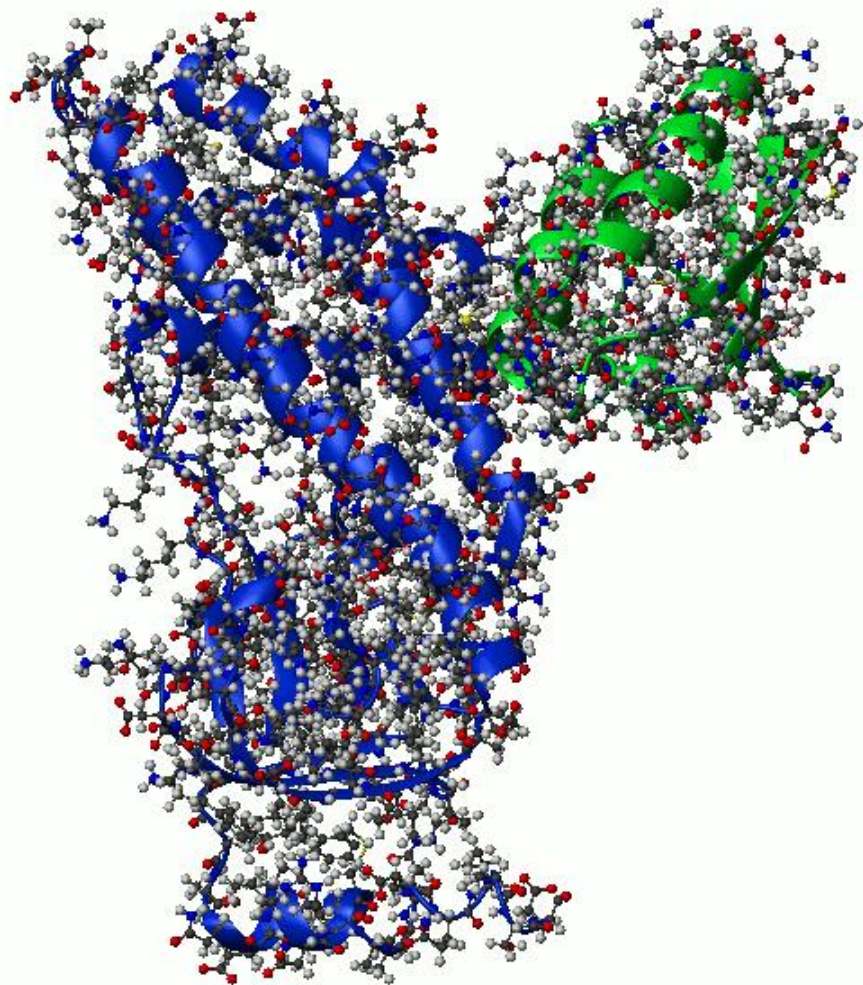
EIN – HPr complex



10 lowest energy structures

## Application to an asymmetric complex

EIN – HPr complex

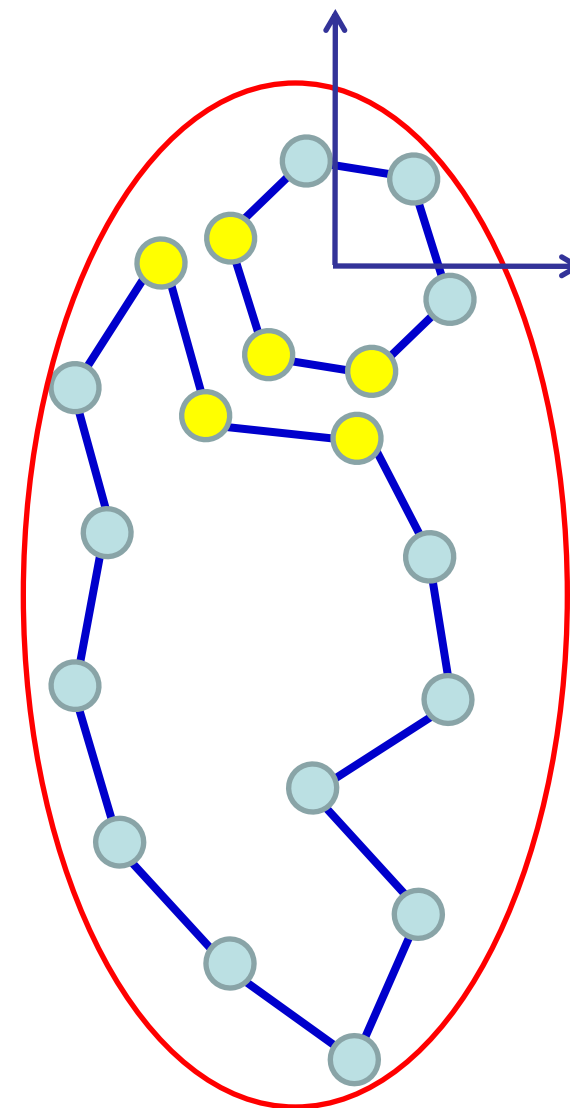
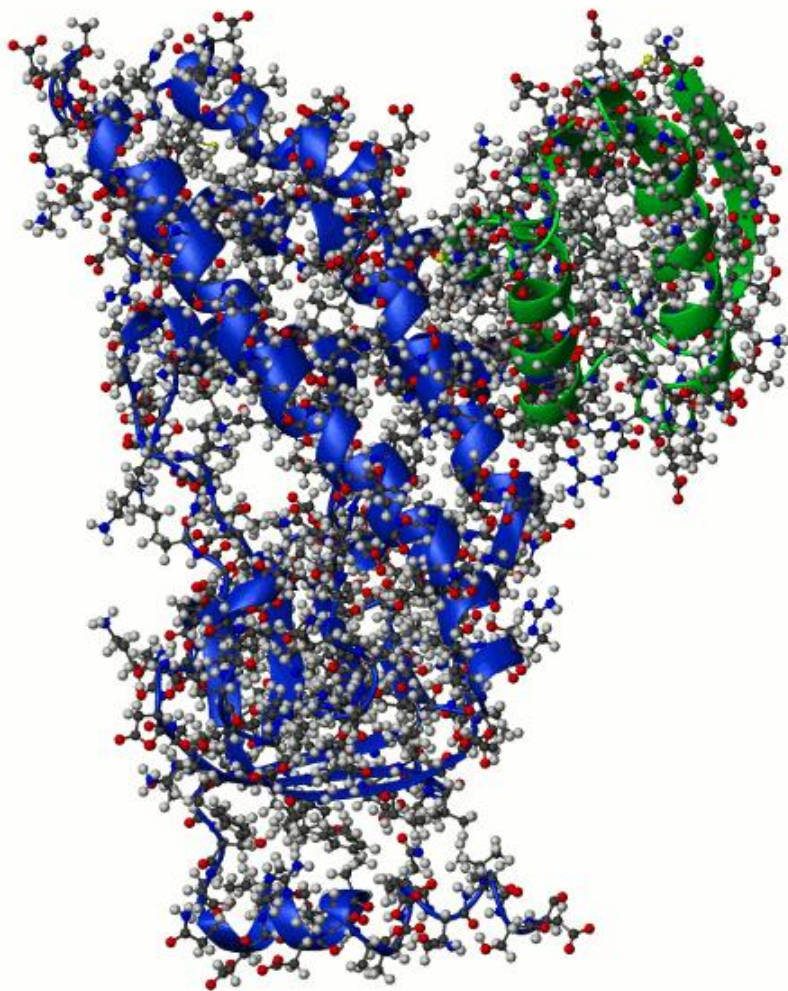


10 lowest energy structures



## Application to an asymmetric complex

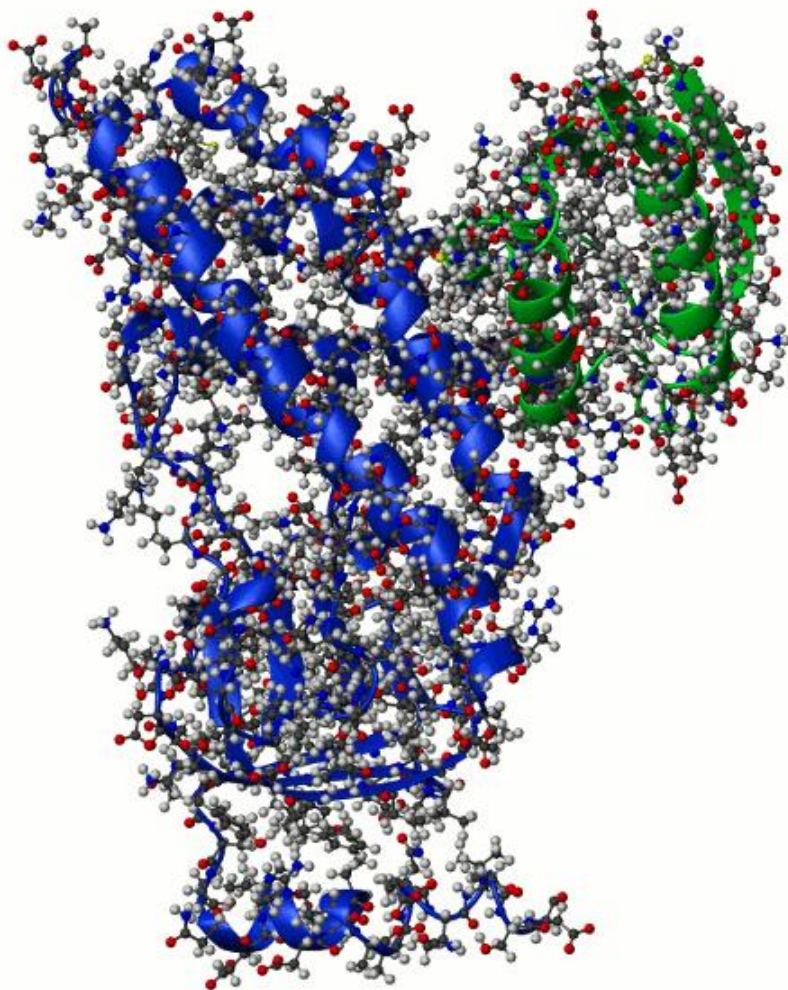
EIN – HPr complex



10 lowest energy structures

## Application to an asymmetric complex

EIN – HPr complex



Randomization of domain positions and Rigid body dynamics repeated 10 times; then the lowest energy structure submitted to final simulated annealing part of the protocol

512 structures calculated.

Experimental restrains were

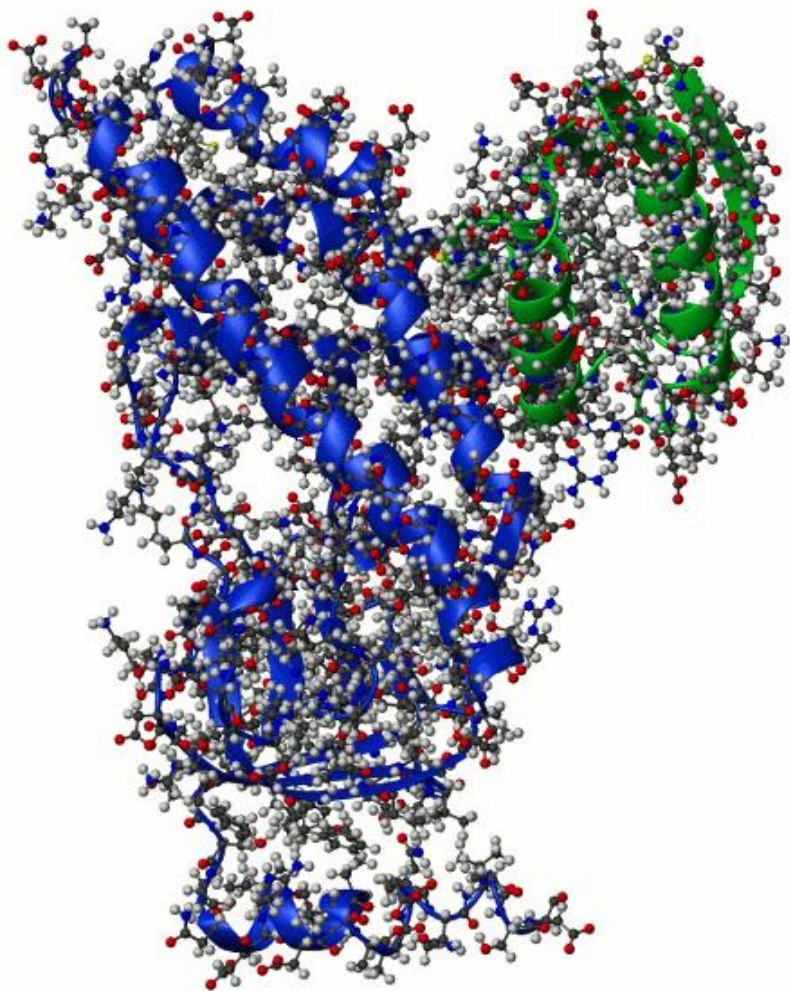
Components of Rotation Diffusion Tensor  
and

Highly ambiguous distance restraints  
from chemical shift perturbation mapping

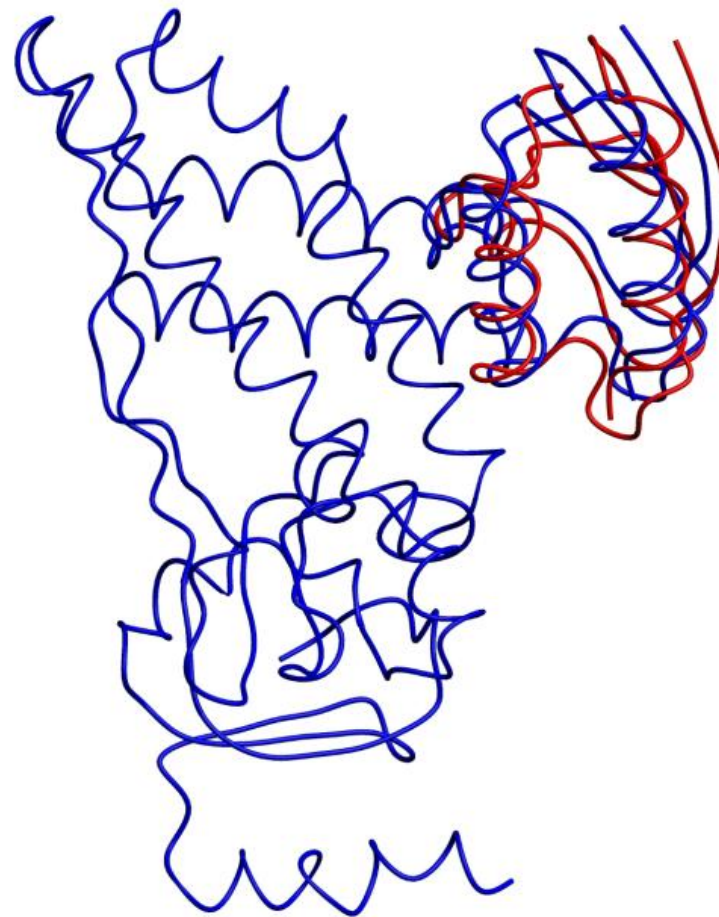
10 lowest energy structures

## Application to an asymmetric complex

EIN – HPr complex



10 lowest energy structures



C $\alpha$  RMSD

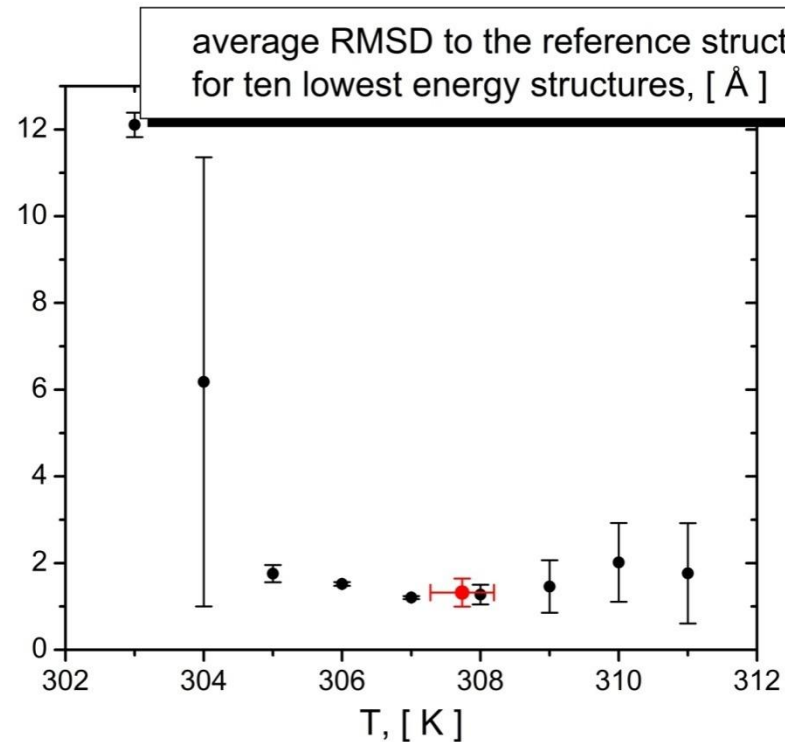
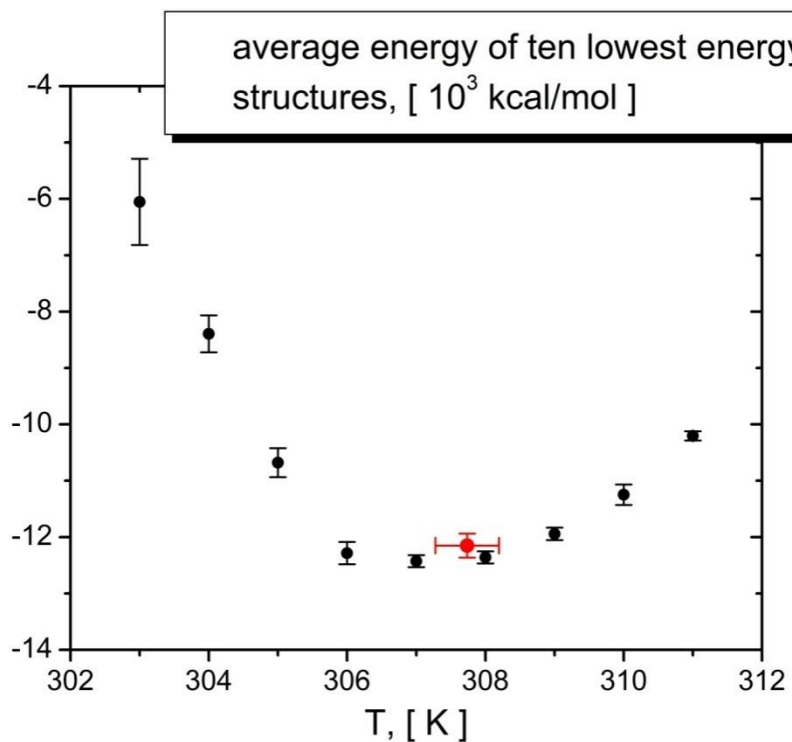
$1.20 \pm 0.03$  [Å]

## Application to an asymmetric complex

Experimental temperature: 313 K

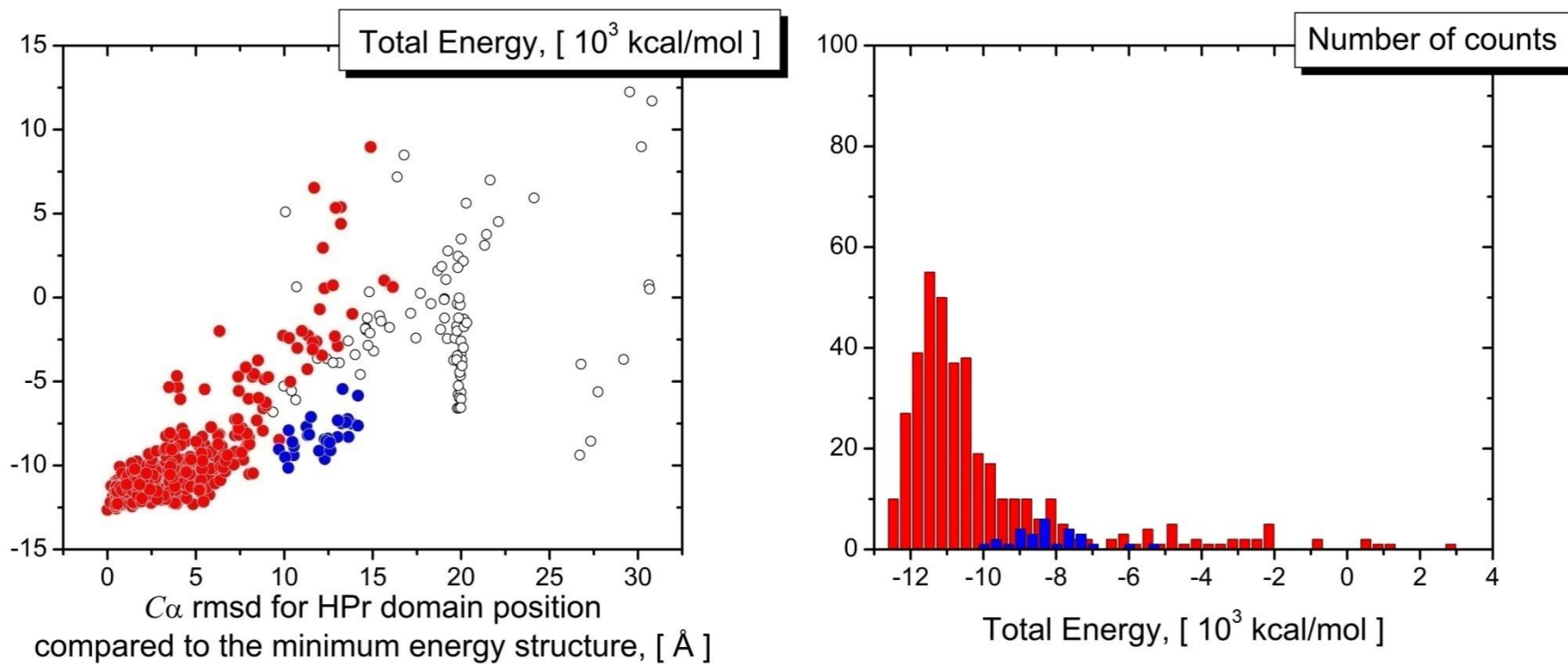
Temperature of the minimum: 307 K

Optimized Temperature:  $307.7 \pm 0.5$



# Application to an asymmetric complex

EIN – HPr complex



75 % in the largest cluster of solutions (red)

## **CONCLUSIONS**

Accuracy of the structures obtained using shape restraints derived from protein rotation tensor is comparable to the accuracy of standard structure determination protocols.

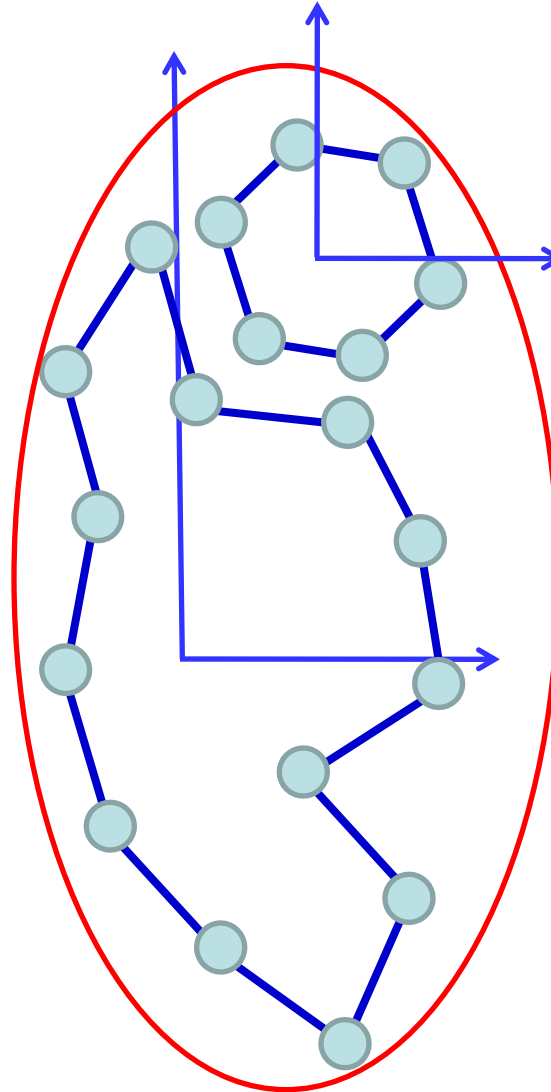
When refining structures of globular proteins, these restraints, in combination with other information, could help to solve the problem of poor packing density of NMR protein structures.

When assembling protein complexes, the relaxation data even for one domain of the complex are enough to drive accurate domain assembly.

In some cases these restraints can be the only experimental information necessary to obtain correct domain assembly.

# Immediate Future directions

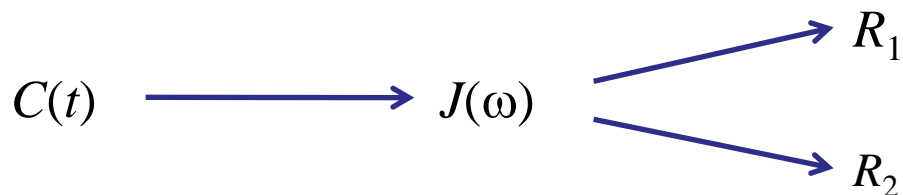
Orientation information from Diffusion Tensor



## Future directions

Structural information from residue specific relaxation NMR rates

Correlation function      Spectral density      Experimentally Measured relaxation rates



Rotation of a protein as a whole

Protein Domain Mobility

$$\begin{aligned}
 C(t) = & \sum_{l,l'=-2}^2 \sum_{m,m'=-2}^2 \sum_{n,n'=-2}^2 \sum_{k,k'=-2}^2 \langle D_{ql}^{2*}(\Omega_{LP}(0)) D_{q'l'}^2(\Omega_{LP}(t)) \rangle \times \\
 & \times \langle D_{lm}^{2*}(\Omega_{PD}(0)) D_{l'm'}^2(\Omega_{PD}(t)) \rangle \times \\
 & \times \langle D_{mn}^{2*}(\Omega_{DR}(0)) D_{m'n'}^2(\Omega_{DR}(t)) \rangle \times \\
 & \times \langle D_{nk}^{2*}(\Omega_{RI}(0)) D_{n'k'}^2(\Omega_{RI}(t)) \rangle
 \end{aligned}$$

Protein structure

Local mobility

The equation is annotated with arrows pointing from descriptive text to specific terms in the product. 'Rotation of a protein as a whole' points to the first term. 'Protein Domain Mobility' points to the second term. 'Protein structure' points to the third term. 'Local mobility' points to the fourth term.



## Future directions

Structural information from residue specific relaxation NMR rates

$R_2/R_1$  Independent of the local mobility

Rotation of a protein as a whole

$$C_{qm,qn}^{LP}(t) = \left\langle D_{q,m}^{(2)*}(\Omega_{L \rightarrow P}^0) D_{q,n}^{(2)}(\Omega_{L \rightarrow P}^t) \right\rangle_{L \rightarrow P} = \frac{1}{5} \sum_{z=-2}^2 e^{-E_z t} a_{z,m}^* a_{z,n}$$

Protein structure

$$C_{ks,lh}^{DR}(t) = \left\langle D_{k,s}^{(2)*}(\Omega_{DR}(0)) D_{l,h}^{(2)}(\Omega_{DR}(t)) \right\rangle_{DR} = D_{k,s}^{(2)*}(\Omega_{DR}) D_{l,h}^{(2)}(\Omega_{DR})$$

**Favro**, *Phys. Rev.* 1960

**Woessner**, *J. Chem. Phys.* 1962

## Future directions

Structural information from residue specific relaxation NMR rates

$R_1$  and  $R_2$  separate

Rotation of a protein as a whole

$$C_{qm,qn}^{LP}(t) = \left\langle D_{q,m}^{(2)*}(\Omega_{L \rightarrow P}^0) D_{q,n}^{(2)}(\Omega_{L \rightarrow P}^t) \right\rangle_{L \rightarrow P} = \frac{1}{5} \sum_{z=-2}^2 e^{-E_z t} a_{z,m}^* a_{z,n}$$

Protein structure

$$C_{ks,lh}^{DR}(t) = \left\langle D_{k,s}^{(2)*}(\Omega_{DR}(0)) D_{l,h}^{(2)}(\Omega_{DR}(t)) \right\rangle_{DR} = D_{k,s}^{(2)*}(\Omega_{DR}) D_{l,h}^{(2)}(\Omega_{DR})$$

Local mobility

$$C_{s_0,h_0}^{RI}(t) = \delta_{s,0} \delta_{h,0} \left[ S^2 + (1 - S^2) \exp\{-t/\tau_l\} \right]$$

Requires ensemble refinement  
to evaluate order parameters  $S^2$

**Favro**, *Phys. Rev.* 1960

**Woessner**, *J. Chem. Phys.* 1962

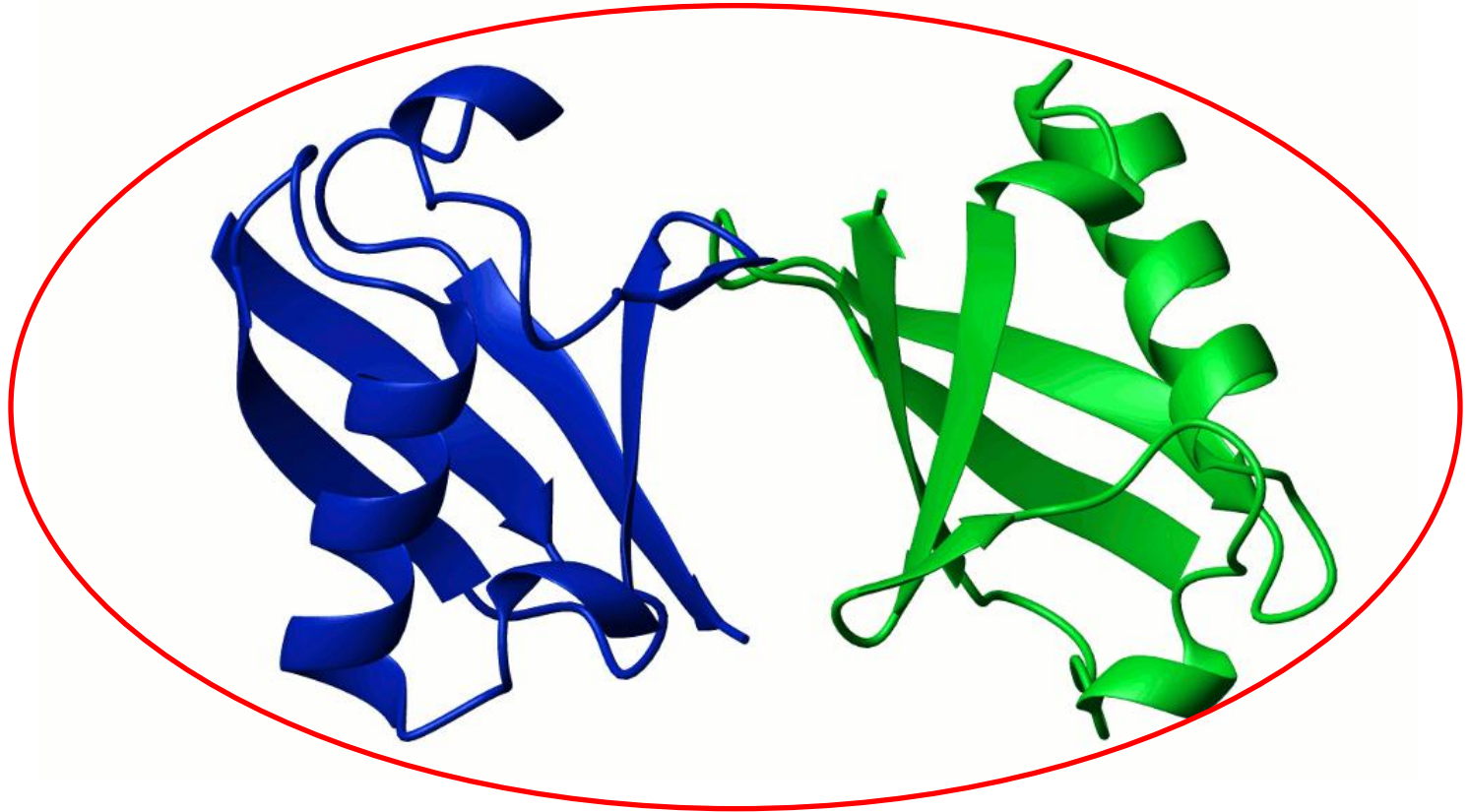
**Lipari & Szabo**

*J. Am. Chem. Soc.* 1982

## Far Future directions

Structural information from residue specific relaxation NMR rates

Protein Domain Mobility



## ACKNOWLEDGMENTS

Co-authors of the paper:

Jeong-Yong Suh,  
Alexander Grishaev,  
G. Marius Clore,  
Charles D. Schwieters

For the help  
with the NMR data on EIN

Daniel Garrett

For the help  
with Xplor-NIH code

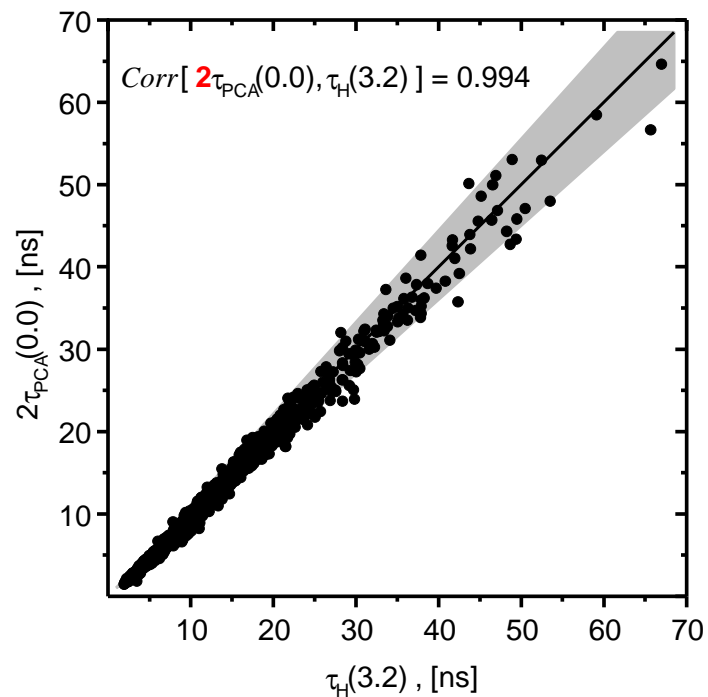
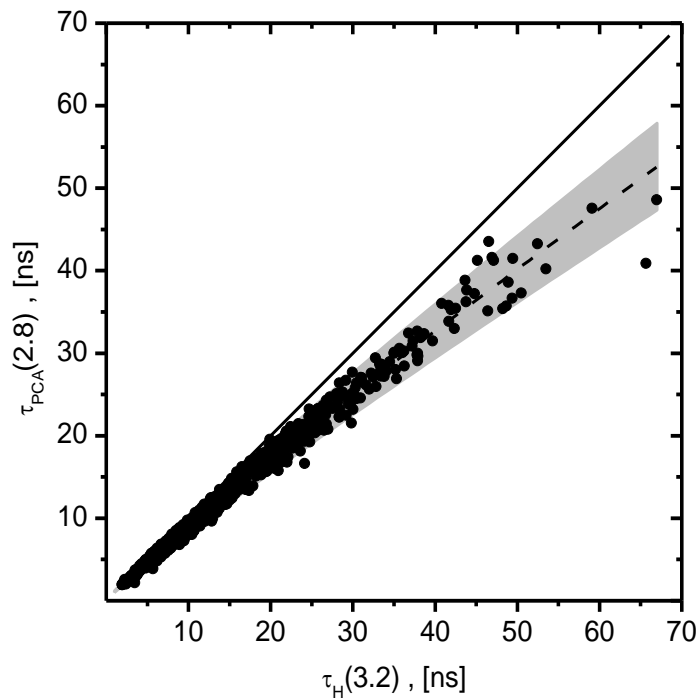
John Kuszewski

For funding

National Research Council

# Correlation times for 841 protein structures

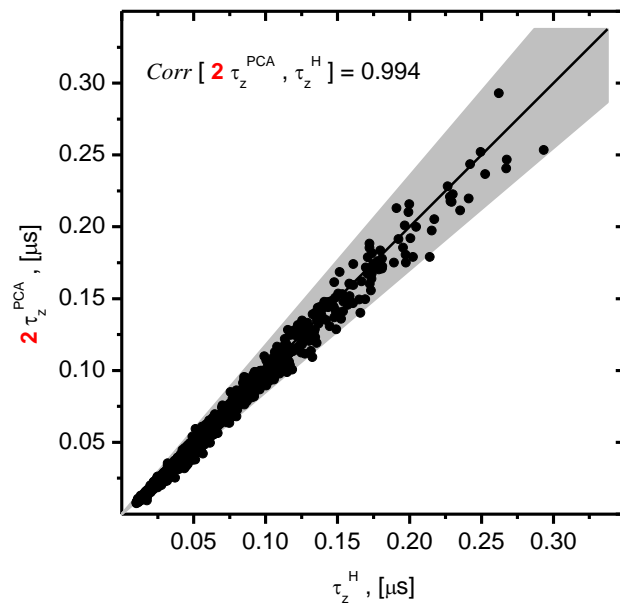
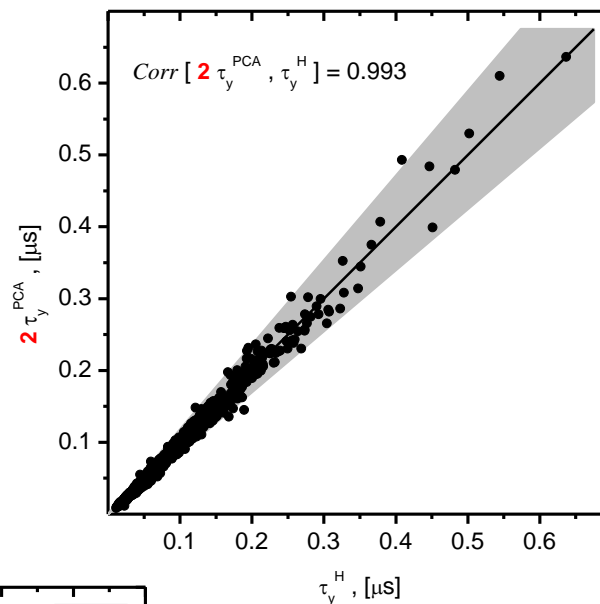
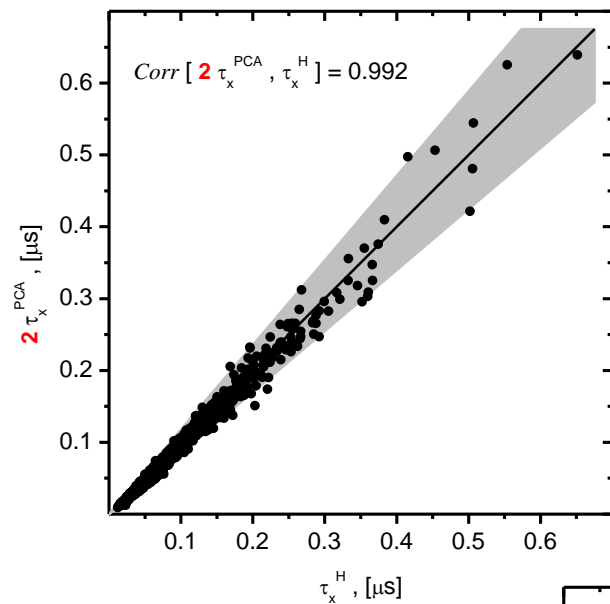
## Hydration shell effect



Power law  $\tau_{PCA} = M\tau_H^q$   $q \sim 0.923$

Fractal surface dimension  $d_f = 2/q \sim 2.2 \div 2.3$

# Correlations for diffusion tensor components



# Correlations of diffusion tensors orientations

